

Mass spectrometry-based proteomics

BMI/CS 776

www.biostat.wisc.edu/bmi776/

Spring 2020

Daifeng Wang

daifeng.wang@wisc.edu

Goals for lecture

Key concepts

- Benefits of mass spectrometry
- Generating mass spectrometry data
- Computational tasks
- Matching spectra and peptides

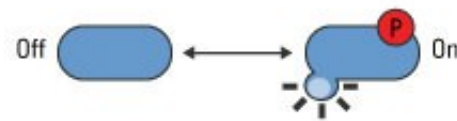
Mass spectrometry uses

- Mass spectrometry is like the protein analog of RNA-seq
 - Quantify abundance or state of all (many) proteins
 - No need to specify proteins to measure in advance
- Other applications in biology
 - Targeted proteomics
 - Metabolomics
 - Lipidomics

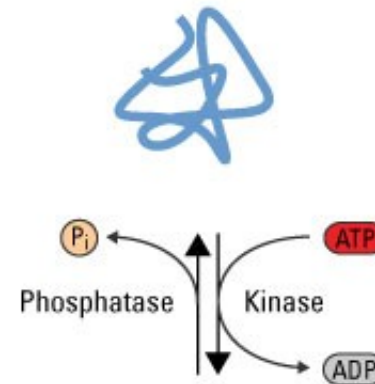
Advantages of proteomics

- Proteins are functional units in a cell
 - Protein abundance directly relevant to activity
- Post-translational modifications
 - Change protein state

Phosphorylation
in signaling



Thermo Fisher Scientific



Histone
modifications

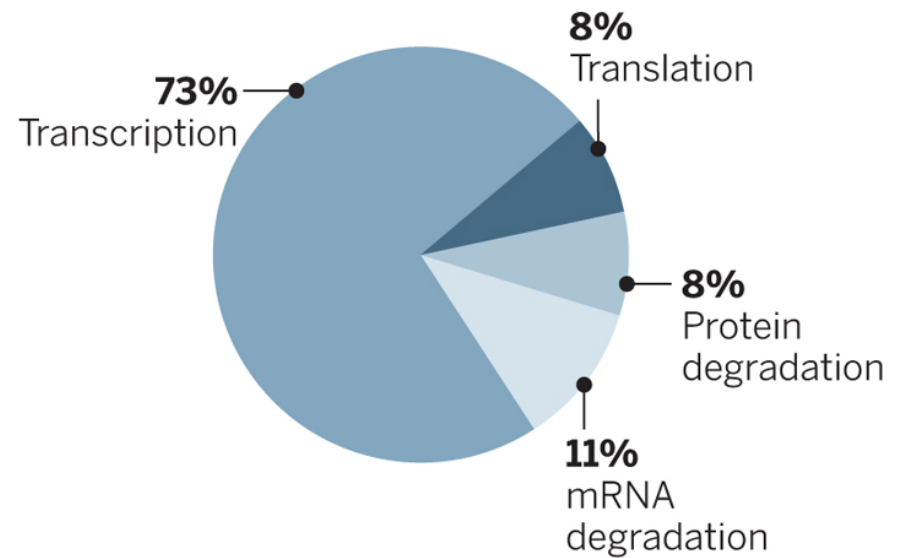


Latham *Nature Structural & Molecular Biology* 2007; Katie Ris-Vicari

Estimating protein levels from gene expression

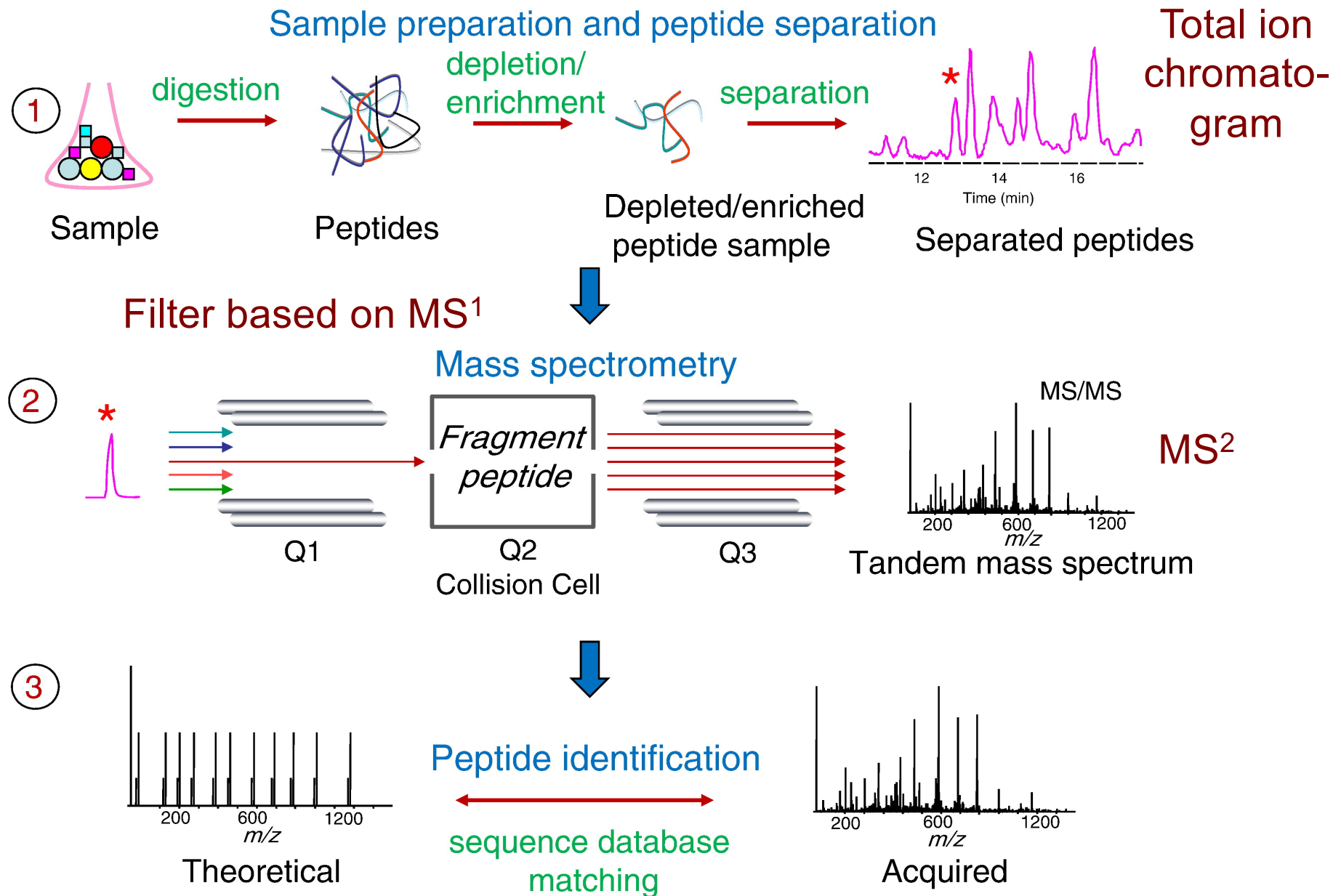
- Correlation between gene expression and protein abundance has been debated
- Gene expression tells us nothing about post-translational modifications

Contribution to protein levels



Li and Biggin *Science* 2015

Mass spectrometry workflow

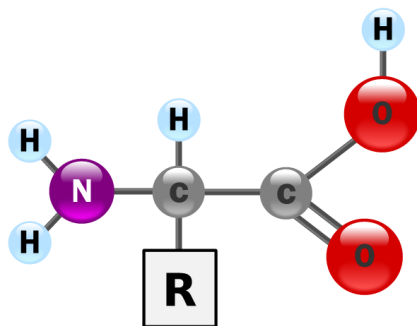


Amino Acids

- 20 amino acids
- Building blocks of proteins
- Known molecular weight
- Common template

Amino-terminal

Carboxy-terminal

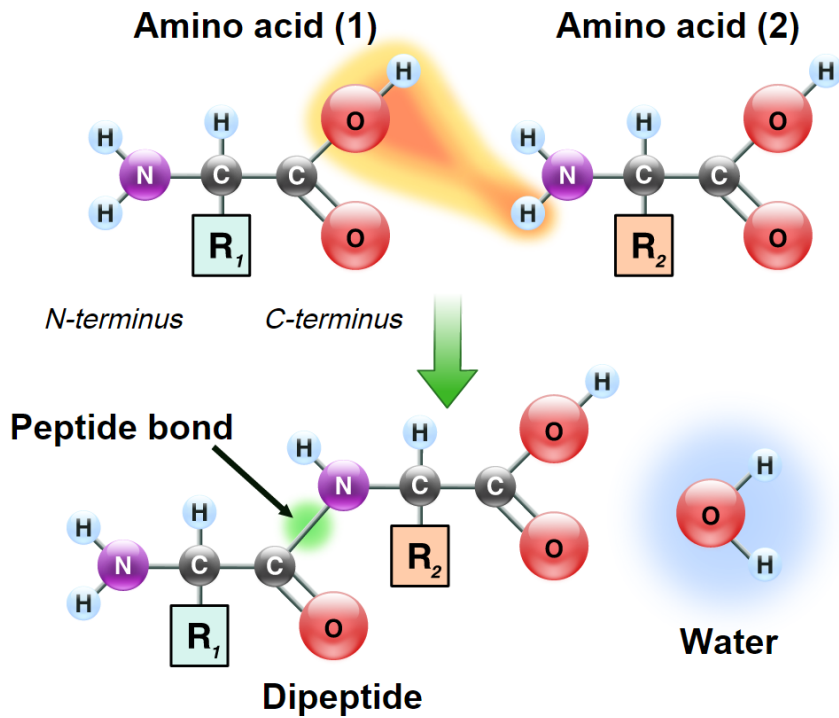


Wikipedia, Yassine Mrabet

NONPOLAR, HYDROPHOBIC		POLAR, UNCHARGED	
	R GROUPS		
Alanine Ala A MW = 89	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_3 \end{array}$	$\begin{array}{c} \text{H} - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Glycine Gly G MW = 75
Valine Val V MW = 117	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}(\text{CH}_3)_2 \end{array}$	$\begin{array}{c} \text{HO-CH}_2 - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Serine Ser S MW = 105
Leucine Leu L MW = 131	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{CH}(\text{CH}_3)_2 \end{array}$	$\begin{array}{c} \text{OH} \\ \\ \text{CH}_3 - \text{CH} - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Threonine Thr T MW = 119
Isoleucine Ile I MW = 131	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}(\text{CH}_3) - \text{CH}_2 - \text{CH}_3 \end{array}$	$\begin{array}{c} \text{HS-CH}_2 - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Cysteine Cys C MW = 121
Phenylalanine Phe F MW = 131	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{C}_6\text{H}_5 \end{array}$	$\begin{array}{c} \text{HO-C}_6\text{H}_4 - \text{CH}_2 - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Tyrosine Tyr Y MW = 181
Tryptophan Trp W MW = 204	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{C}_8\text{H}_6\text{N}_2 \end{array}$	$\begin{array}{c} \text{NH}_2 \\ \\ \text{O}=\text{C} - \text{CH}_2 - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Asparagine Asp N MW = 132
Methionine Met M MW = 149	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{CH}_2 - \text{S} - \text{CH}_3 \end{array}$	$\begin{array}{c} \text{NH}_2 \\ \\ \text{O}=\text{C} - \text{CH}_2 - \text{CH}_2 - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Glutamine Gln Q MW = 146
Proline Pro P MW = 115	$\begin{array}{c} \text{OOC}^- \\ \\ \text{CH} - \text{CH}_2 - \text{CH}_2 - \text{CH}_2 - \text{NH} \end{array}$	POLAR BASIC $\begin{array}{c} \text{NH}_3^+ - \text{CH}_2 - (\text{CH}_2)_3 - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Lysine Lys K MW = 146
Aspartic acid Asp D MW = 133	POLAR ACIDIC $\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{C}(=\text{O})\text{O}^- \end{array}$	$\begin{array}{c} \text{NH}_2 \\ \\ \text{N H}_2^+ = \text{C} - \text{NH} - (\text{CH}_2)_3 - \text{CH} - \text{COO}^- \\ \\ \text{N H}_3^+ \end{array}$	Arginine Arg R MW = 174
Glutamine acid Glu E MW = 147	$\begin{array}{c} \text{OOC}^- \\ \\ \text{H}_3\text{N}^+ - \text{CH} - \text{CH}_2 - \text{CH}_2 - \text{C}(=\text{O})\text{O}^- \end{array}$	$\begin{array}{c} \text{HN}=\text{C} - \text{CH}_2 - \text{CH} - \text{COO}^- \\ \\ \text{H N}^+ \end{array}$	Histidine His H MW = 155

Peptide fragmentation

Peptide bond

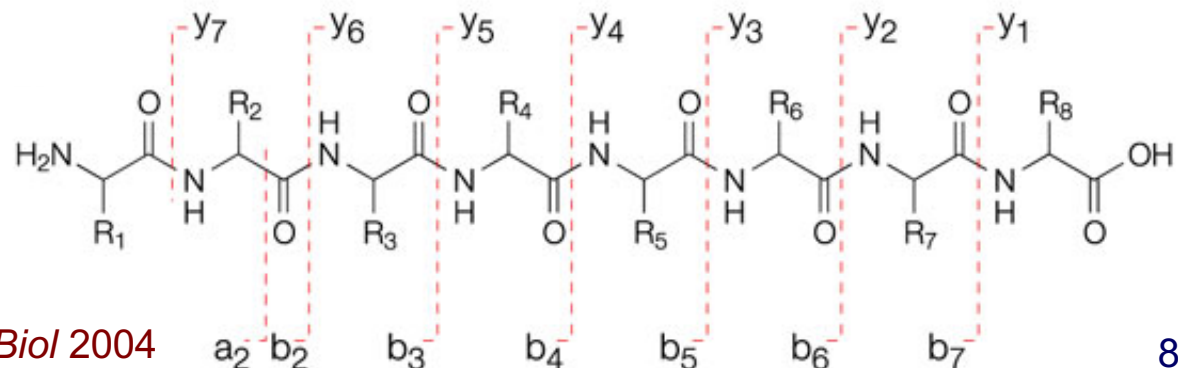


Wikipedia, Yassine Mrabet

- Select similar peptides from MS¹
- Fragment with high energy collisions
- Break peptide bonds

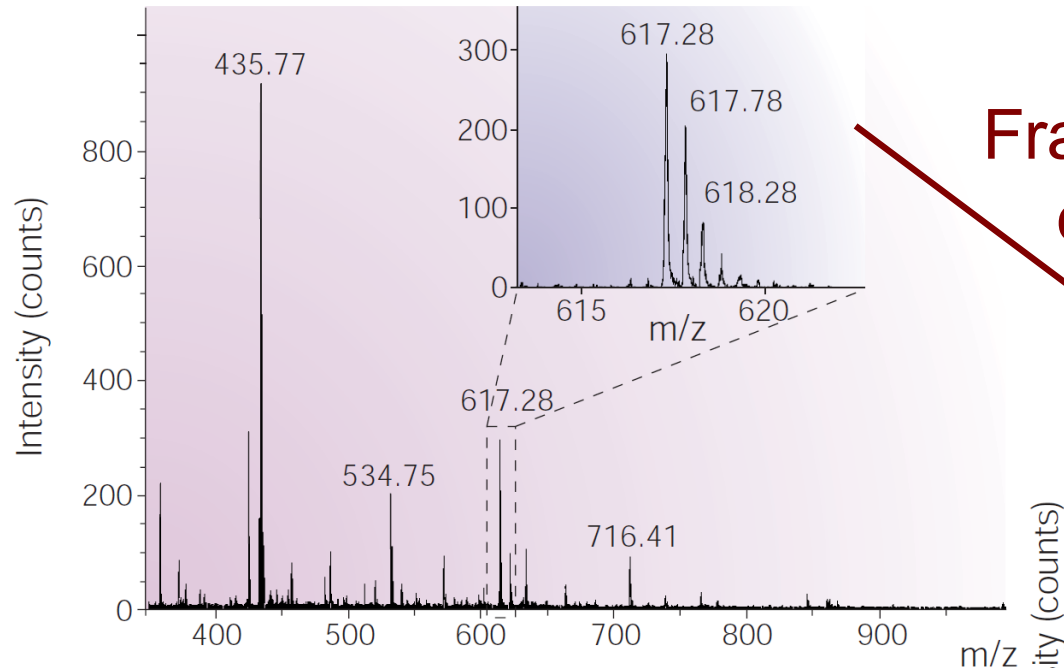
Charge on amino-terminal (b) or carboxy-terminal fragment (y)

Subscript = # R groups retained



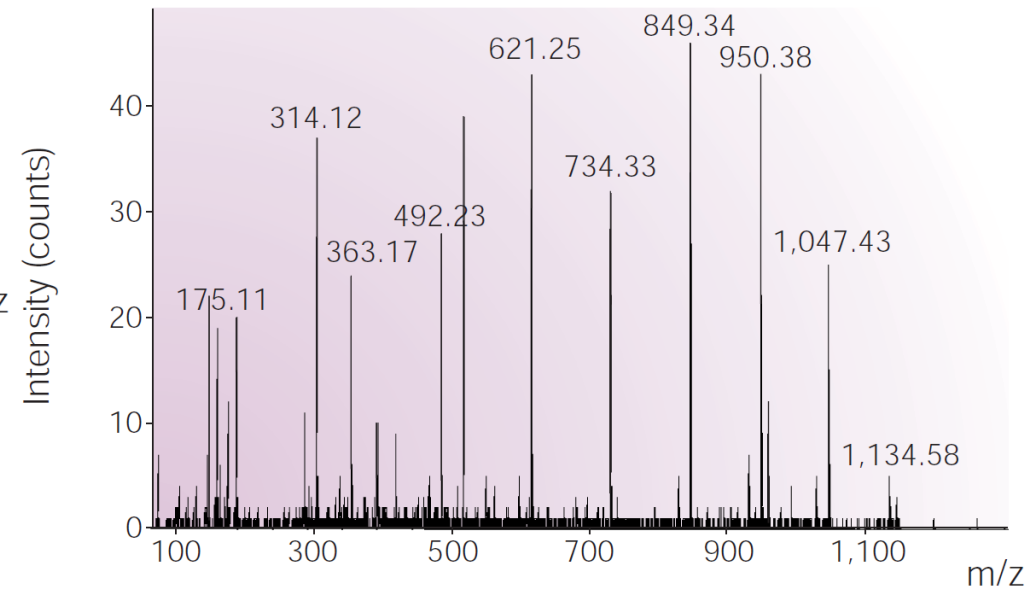
Mass spectra

MS¹



Fragment and analyze
one precursor ion

MS²



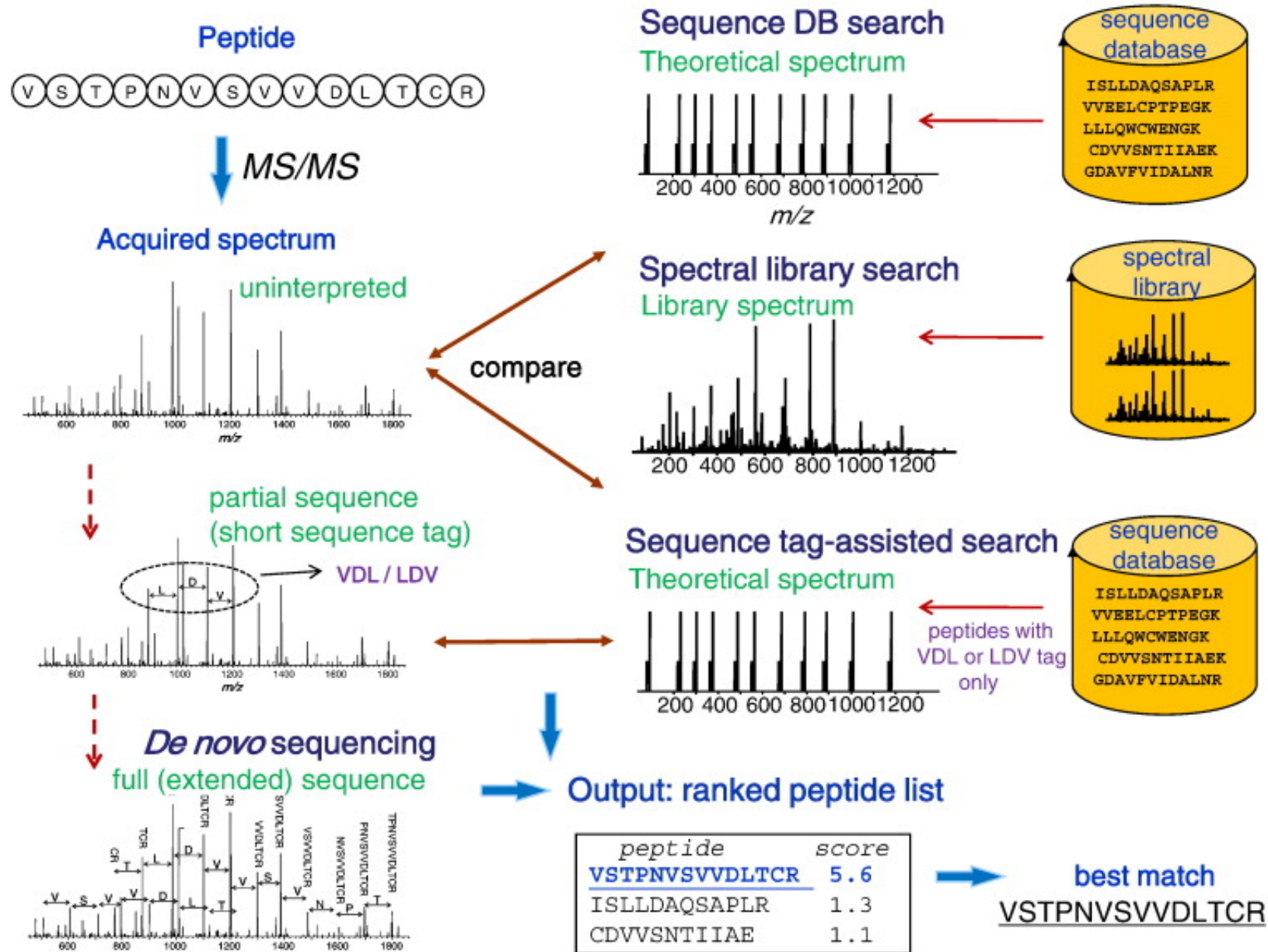
Steen and Mann *Nat Rev Mol Cell Biol* 2004

Mass-to-charge ratio



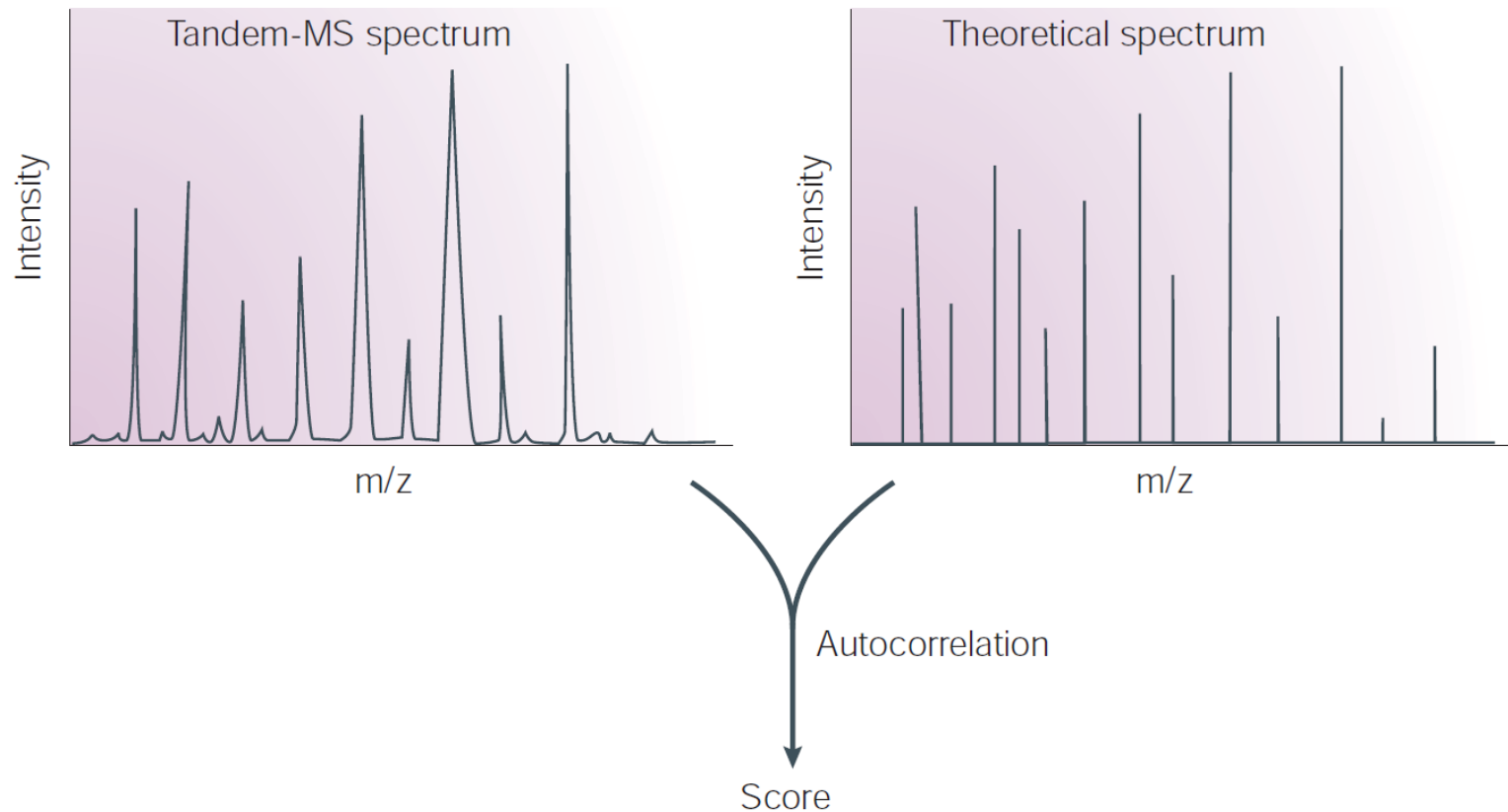
Spectrum contains information about amino
acid sequence, fragment at different bonds

From spectra to peptides



Sequence database search

- Need to define a scoring function
- Identify peptide-spectrum match (PSM)



SEQUEST

- Cross correlation (xcorr)
- Similarity between theoretical spectrum (x) and acquired spectrum (y)
- Correction for mean similarity at different offsets

$$\text{xcorr} = R_0 - \left(\sum_{\tau=-75}^{\tau=+75} R_{\tau} \right) / 151$$

Actual similarity

$$R_{\tau} = \sum x[i] \cdot y[i + \tau]$$

Theoretical

Acquired

Offsets

Fast SEQUEST

- SEQUEST originally only applied to top 500 peptides based on coarse filtering score

$$\text{xcorr} = x_0 \cdot y_0 - \left(\sum_{\tau=-75}^{\tau=+75} x_0 \cdot y_{\tau} \right) / 151$$

$$\text{xcorr} = x_0 \cdot \left(y_0 - \left(\sum_{\tau=-75}^{\tau=+75} y_{\tau} \right) / 151 \right)$$

$$\text{xcorr} = x_0 \cdot y' \quad \text{where} \quad y' = y_0 - \left(\sum_{\tau=-75, \tau \neq 0}^{\tau=+75} y_{\tau} \right) / 150$$

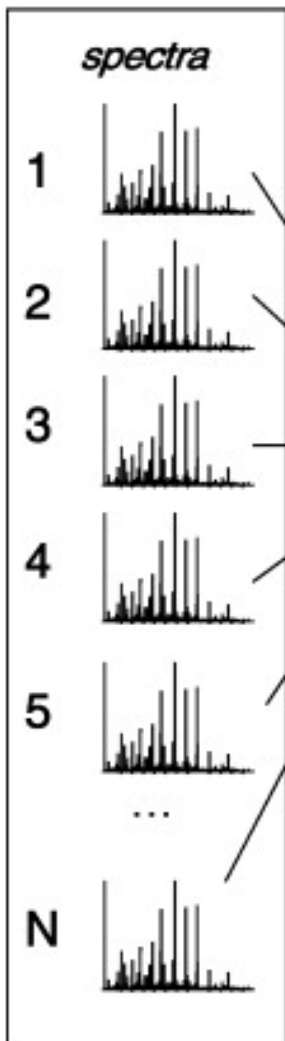
↑ Skip the 0 offset

PSM significance

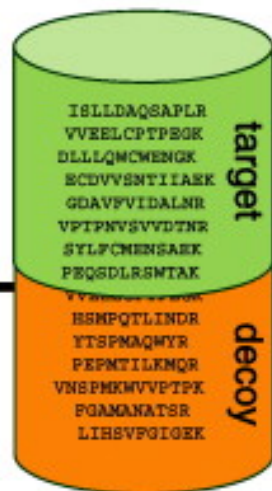
- E-value: expected number of null peptides with score $\geq$ observed score
- Compute FDR from E-value distribution
- Add decoy peptides to database
 - Reversed peptide sequences
 - Used to estimate false discoveries

Target-decoy strategy

Entire dataset, N spectra



database
search



Best match for each spectrum

spec	peptide	score	label
1	ISLLDAQSAPLR	4.5	target
2	VVEELCTPEGK	3.9	target
5	GDAVFVIDALNR	3.6	target
3	VNSPMKWVPTPK	1.7	decoy
4	ECDVVSNTIIAEK	1.5	target
	...		
N	LIHSVFGIGEK	1.1	decoy

(sorted by score)

Nesvizhskii *Journal of Proteomics* 2010

Filtering using
target-decoy
strategy

Apply score threshold S_T

Calculate $N_t(S_T)$ and $N_d(S_T)$:
number of target/decoy PSM with $S \geq S_T$

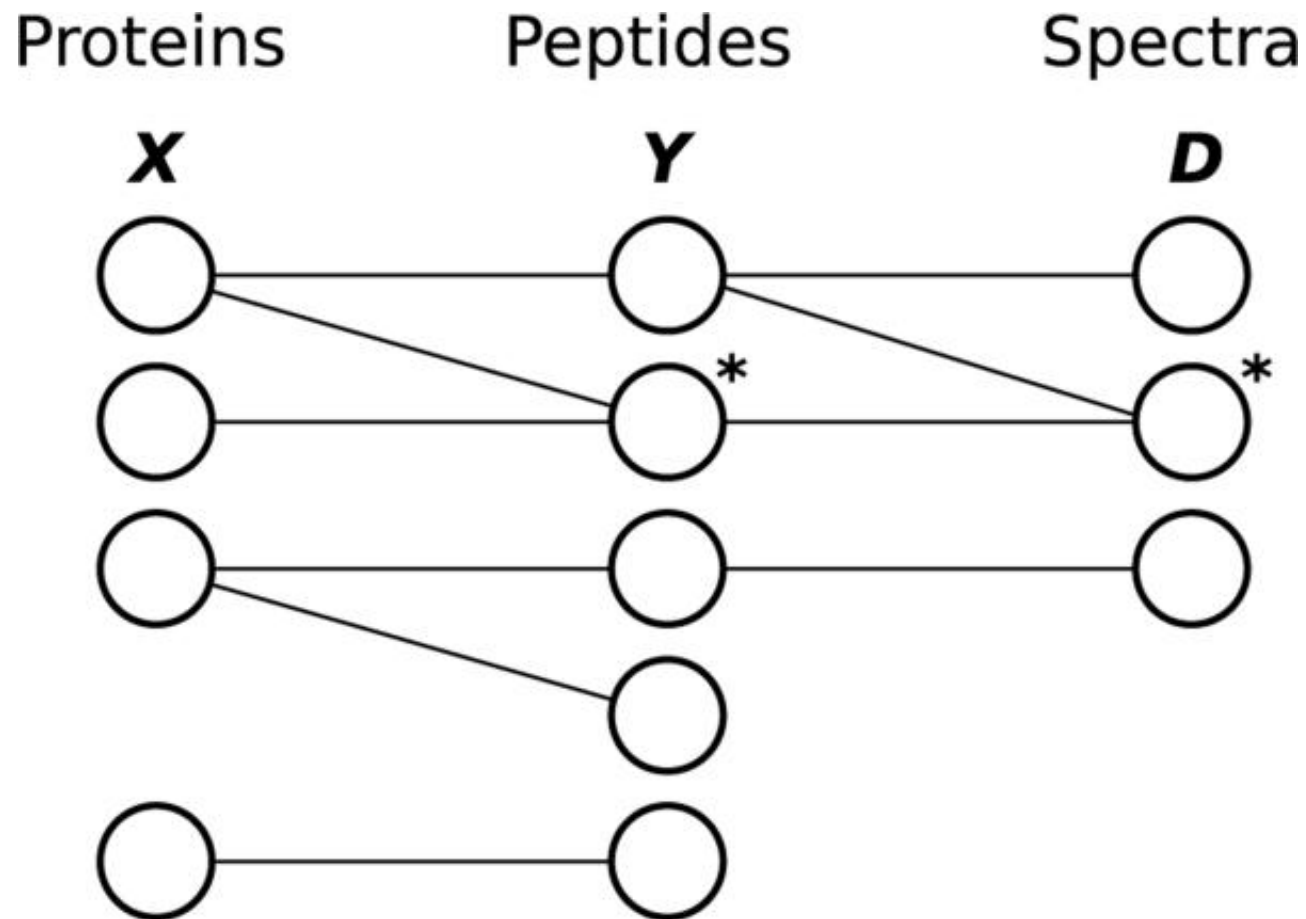
Estimate FDR
$$FDR(S_T) = \frac{N_d(S_T)}{N_t(S_T)}$$

Select threshold S_T to achieve desired FDR

target PSMs above score threshold = $N_t(S_T)$
decoy PSMs above score threshold = $N_d(S_T)$

Identifying proteins

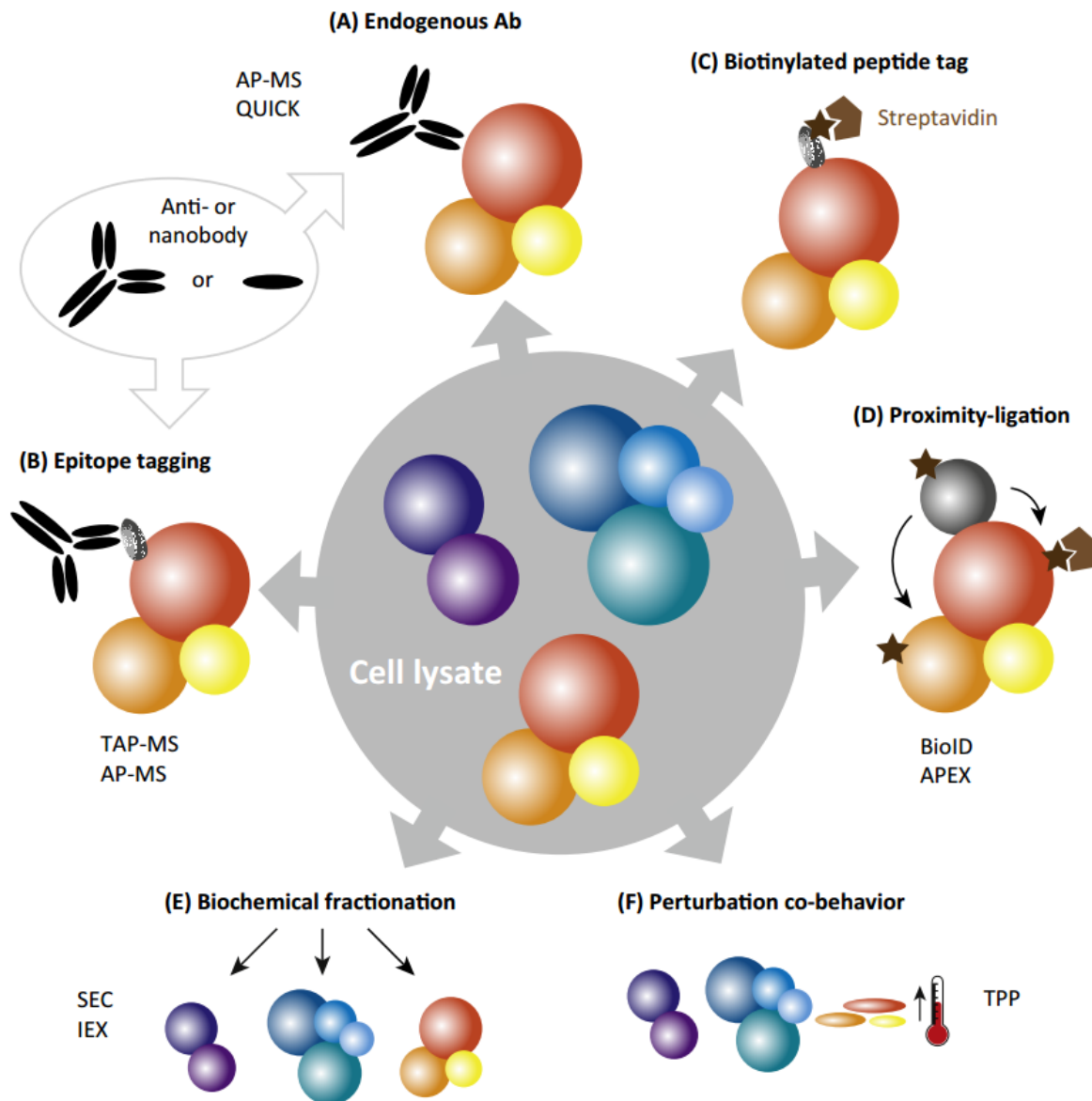
- Even after identifying PSM, still need to identify protein of origin



Mass spectrometry versus RNA-seq

- RNA-seq
 - Transcript → RNA fragment → paired-end read
- Mass spectrometry
 - Protein → peptides → ions → spectrum
- Mapping spectra to proteins more ambiguous than mapping reads to genes
- Spectra state space is enormous

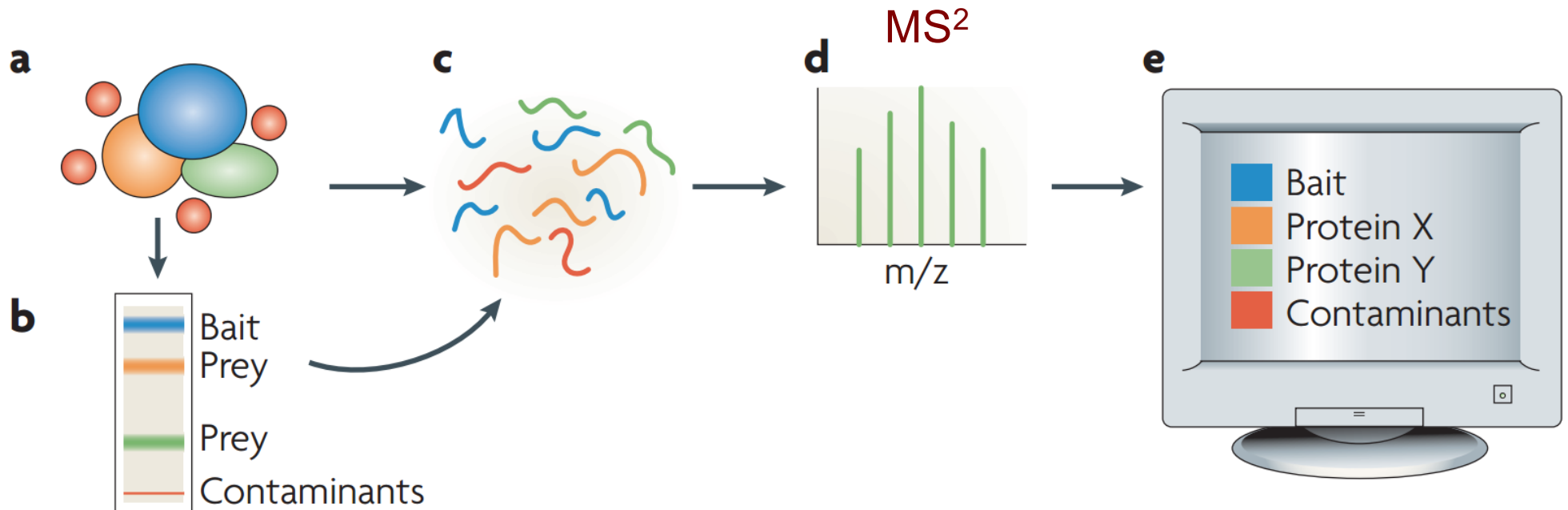
Protein-protein interactions



- Affinity-purification mass spectrometry
- Purify protein of interest, identify complex members

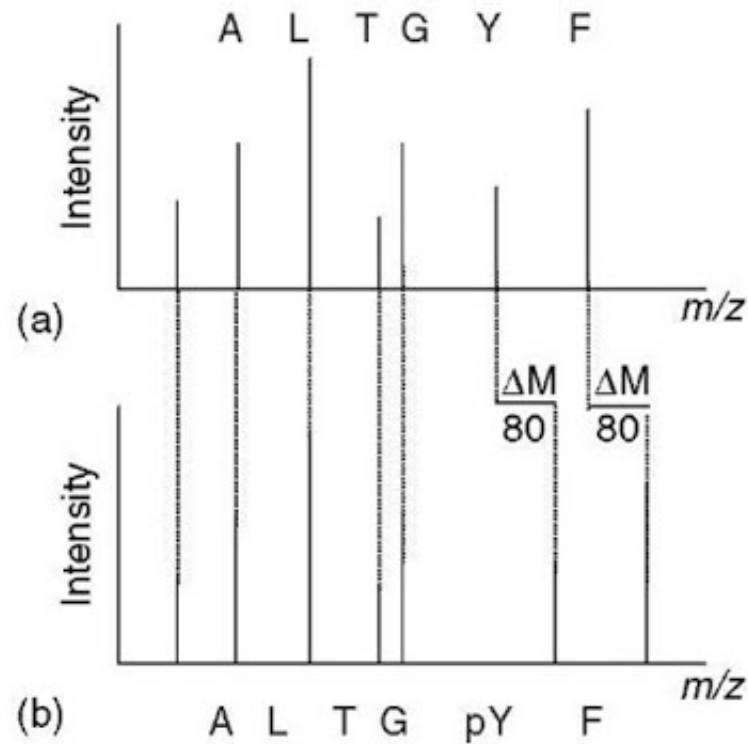
Protein-protein interactions

- Mass spectrometry identifies proteins in the complex
- Must control for contaminants



Post-translational modifications (PTMs)

- Shift the peptide mass by a known quantity



what-when-how

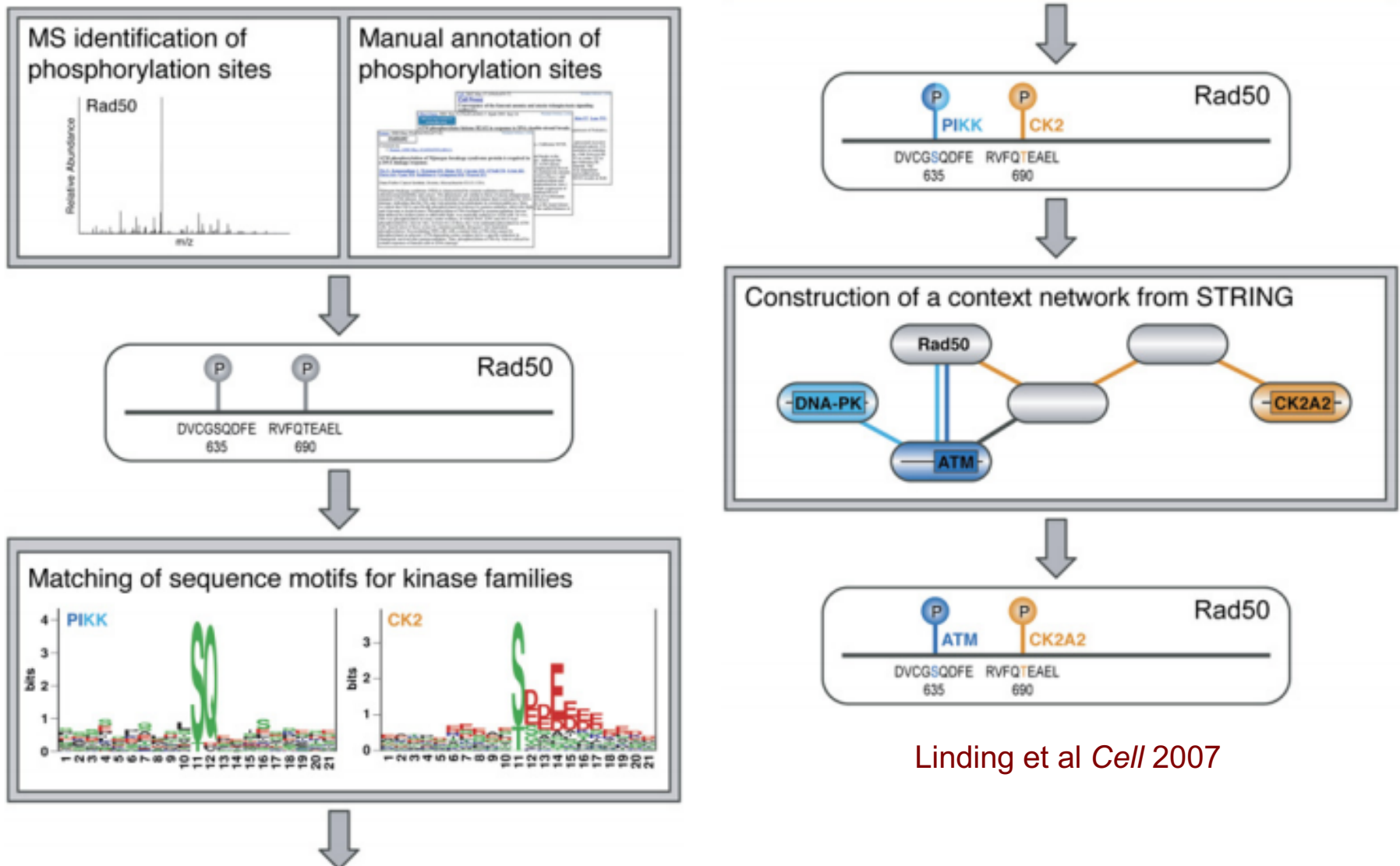
Phosphoproteomics example

Gene	Modified Site	Peptide	Phosphorylation (Treatment / Control)
AGRN	S671	AGPC[160.03]EQAEC[160.03]GS[167.00]GGSGSGEDGDC[160.03]EQELC[160.03]R	4.54
ADAMTS10	S74	RGTGATAES[167.00]R	0.30
CABYR	T16	T[181.01]LLEGISR	0.37
TTC7B	T152	VIEQDET[181.01]R	5.97
STAT3	Y705	K.n[305.21]YC[160.03]RPESQEHPEADPGSAAPY[243.03]LK[432.30].T	4.50

Sychev et al *PLoS Pathogens* 2017

Phosphoproteomics interpretation

- Predict kinases/phosphatases for phospho sites



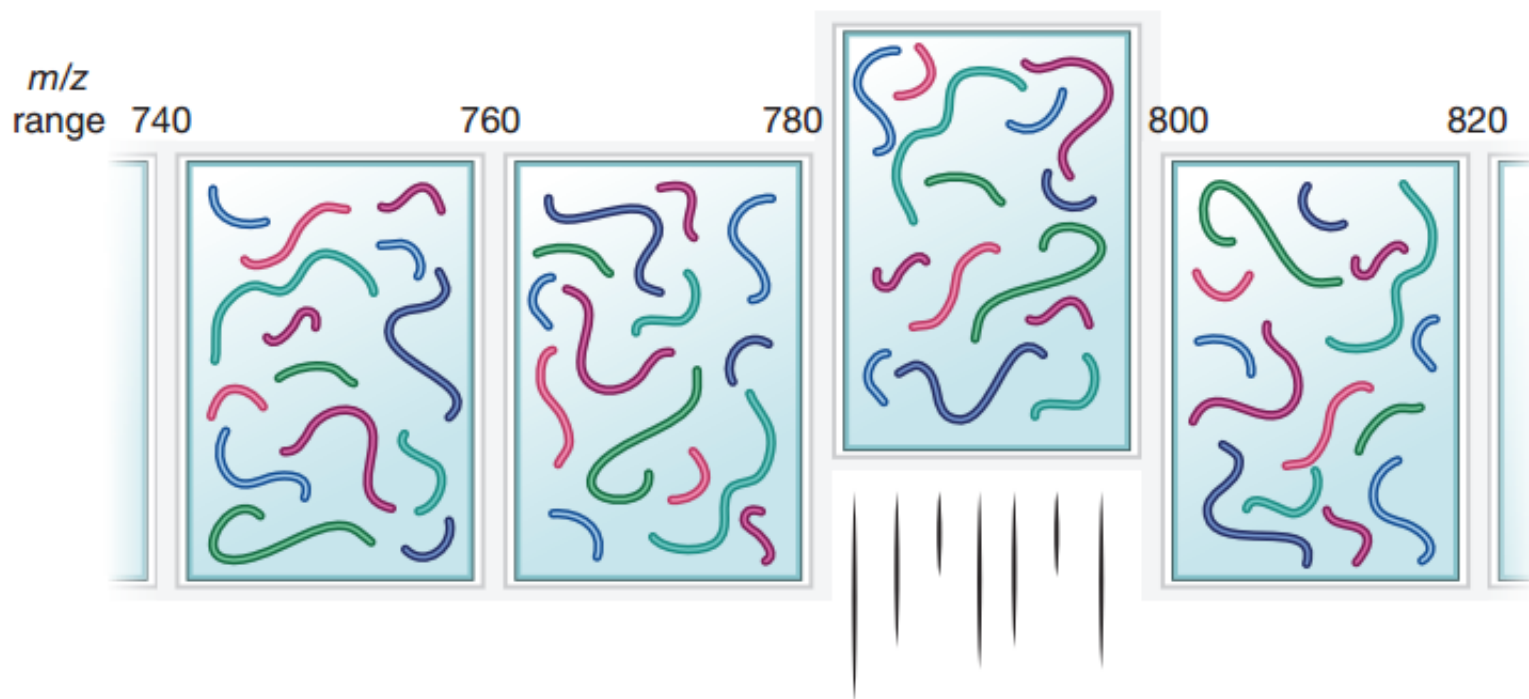
Linding et al *Cell* 2007

Mass spectrometry replicates

- Doesn't identify all proteins in the sample
 - Data dependent acquisition has low overlap across replicates
 - Partly due to biological variation
 - New protocols to overcome this
- Phosphorylation PTMs are especially variable
 - Grimsrud et al *Cell Metabolism* 2012
 - 5 biological replicates
 - 9,558 phosphoproteins identified
 - 5.6% in all replicates

Data independent acquisition

- Not dependent on most abundance signals in MS¹
- Sliding m/z window



Doerr *Nature Methods* 2015

Mass spectrometry summary

- Incredibly powerful for looking at biological processes beyond gene expression
 - Protein abundance
 - Post-translational modifications
 - Metabolites
 - Protein-protein interactions
- Typically reports relative abundance
- Labeling strategies for comparative analysis
 - Compare relative abundance in multiple conditions
- Missing data was a big problem, but improving
- Fully probabilistic analysis pipelines are not the most popular tools
 - Arguably greater diversity in software than RNA-seq