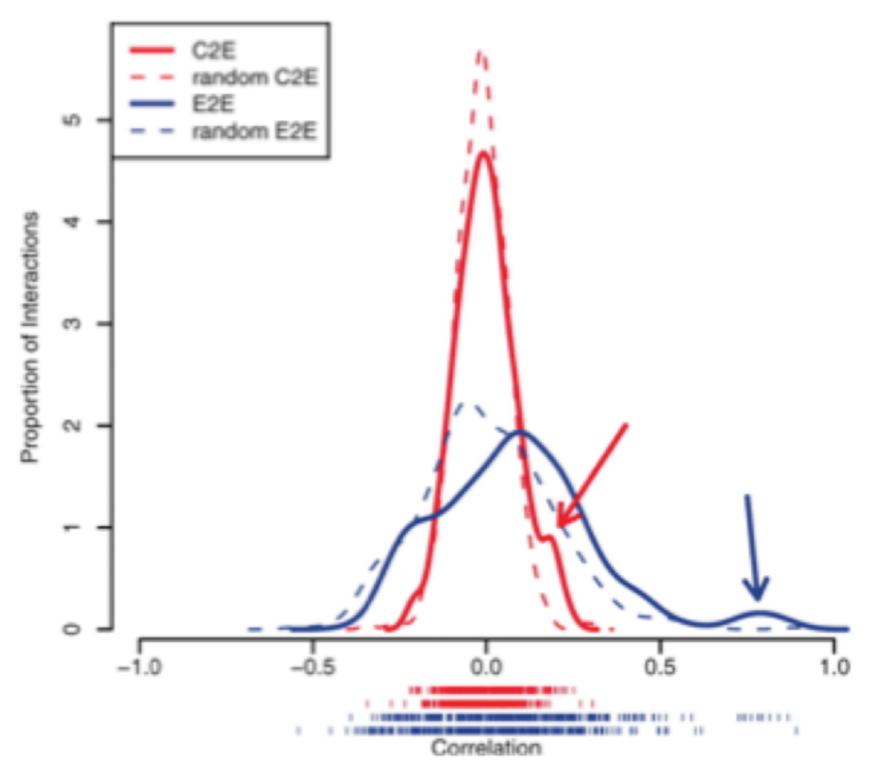
# PARADIGM

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#### NCI Pathway interactions in TCGA GBM data.



Charles J. Vaske et al. Bioinformatics 2010;26:i237-i245



# PARADIGM Approach

We developed an approach called PARADIGM (PAthway Recognition Algorithm using Data Integration on Genomic Models) to infer the activities of genetic pathways from integrated patient data.

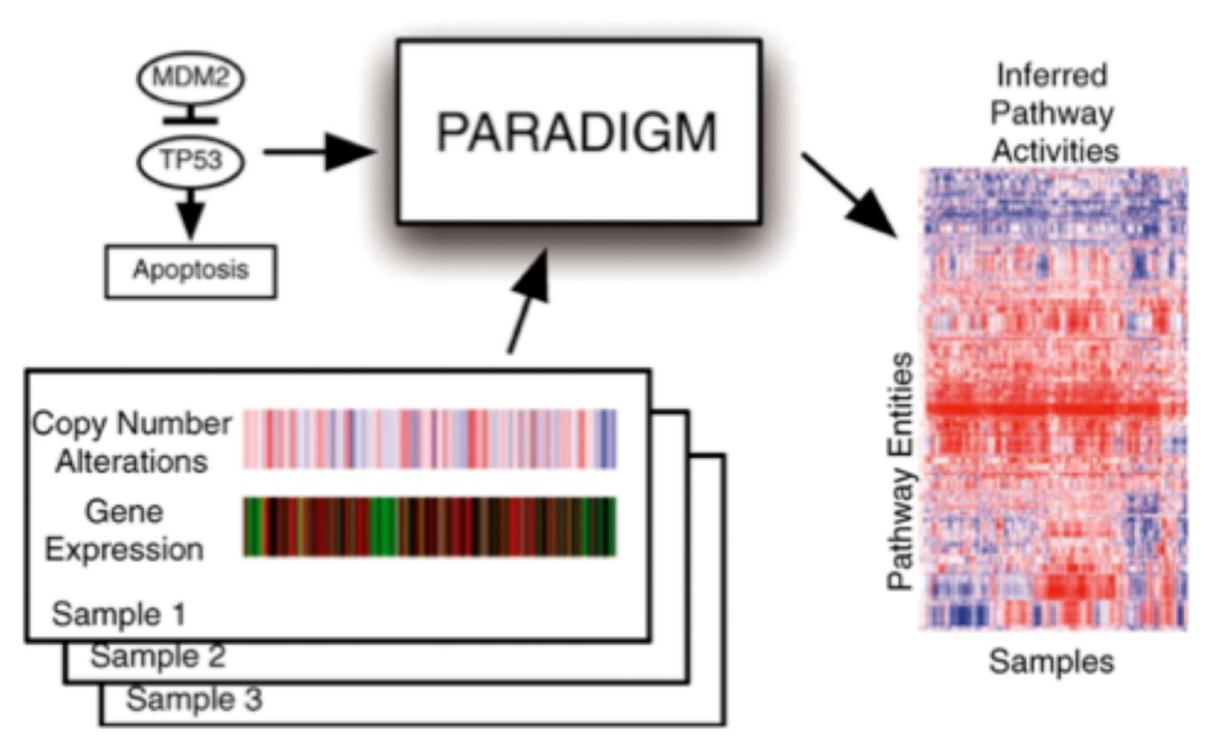
Multiple genome-scale measurements on a single patient sample are combined to infer the activities of genes, products and abstract process inputs and outputs for a single NCI pathway.

# PARADIGM Approach

PARADIGM produces a matrix of integrated pathway activities (IPAs) A where Aij represents the inferred activity of entity i in patient sample j.



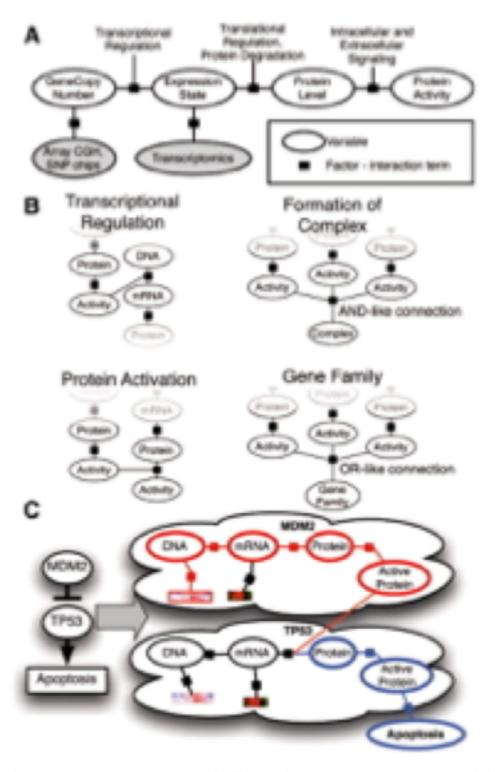
#### Overview of the PARADIGM method.



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#### Conversion of a genetic pathway diagram into a PARADIGM model.



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# Method

### GOAL!

Make a factor graph that represents the underlying pathway.

Each entity can take on one of three states corresponding to activated, nominal or deactivated relative to a control level (e.g. as measured in normal tissue) and encoded as 1, 0 or -1 respectively.

The states may be interpreted differently depending on the type of entity (e.g. gene, protein, etc)

# Factor Graph Goal

The factor graph encodes the state of a cell using a random variable for each entity  $X=\{x1, x2,..., xn\}$  and a set of m non-negative functions, or factors, that constrain the entities to take on biologically meaningful values as functions of one another.

The j-th factor  $\phi$ j defines a probability distribution over a subset of entities x\_j $\subset$ X. The entire graph of entities and factors encodes the joint probability distribution over all of the entities as:

$$P(X) = \frac{1}{Z} \prod_{j=1}^{m} \phi_j(X_j),$$

where  $Z=\prod \sum S \subseteq X$   $\phi$  j(S) is a normalization constant and  $S \subseteq X$  denotes that S is a 'setting' of the variables in X.

### Construction

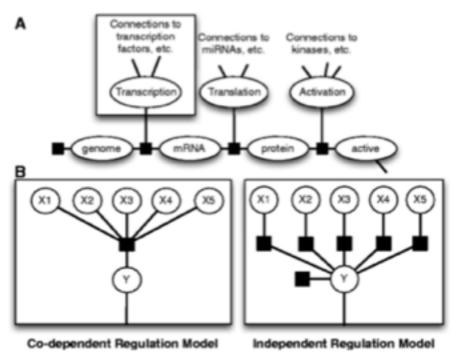
In order to simplify the construction of factors, we first convert the pathway into a directed graph, with each edge in the graph labeled with either positive or negative influence.

Every interaction in the pathway is converted to a single edge in the directed graph.

Using this directed graph, we then construct a list of factors to specify the factor graph.

For every variable xi, we add a single factor  $\phi(Xi)$ , where  $Xi=\{xi\}\cup\{Parents(xi)\}$  and Parents(xi) refers to all the parents of xi in the directed graph.

Factor graph structures in PARADIGM. (A) Central dogma structure shared by all protein coding genes.



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# Filling Out the FG

The expected value was set to the majority vote of the parent variables.

$$\phi_i(x_i, \operatorname{Parents}(x_i)) = \begin{cases} 1 - \epsilon & x_i \text{ is the expected state from } \operatorname{Parents}(x_i) \\ \frac{\epsilon}{2} & \text{otherwise.} \end{cases}$$

If a parent is connected by a positive edge it contributes a vote of +1 times its own state to the value of the factor. (negative edge, then -1)

The variables connected to xi by an edge labeled 'minimum' get a single vote, and that vote's value is the minimum value of these variables, creating an AND-like connection. Similarly the variables connected to xi by an edge labeled 'maximum' get a single vote, and that vote's value is the maximum value of these variables, creating an OR-like connection.

Votes of zero are treated as abstained votes. If there are no votes the expected state is zero.

Otherwise, the majority vote is the expected state, and a tie between 1 and -1 results in an expected state of -1 to give more importance to repressors and deletions.

### Inference

Given patient data, we would like to estimate whether a particular hidden entity xi is likely to be in state a.

For example, how likely TP53's protein activity is -1 (inactivated) or 'Apoptosis' is+1 (activated).

To do this, we first compute the prior probability of the event prior to observing the patient's data.

If Ai(a) represents the singleton assignment set {xi=a} and Φ is the fully specified factor graph, this prior probability is:

$$P(x_i = a | \Phi) = \frac{1}{Z} \prod_{j=1}^{m} \sum_{\mathbf{S} \subseteq A_i(a)} \chi_j \phi_j(\mathbf{S}),$$

### Inference Cont.

The probability that xi is in state a along with all of the observations made for the patient is:

$$P(x_i = a, D | \Phi) = \frac{1}{Z} \prod_{j=1}^{m} \sum_{\mathbf{S} \sqsubseteq_{A_i(a) \cup D} X_j} \phi_j(\mathbf{S}).$$

For the majority of pathways, we use the junction tree inference algorithm with HUGIN updates to infer the probabilities in equations. For pathways that take longer than 3 s of inference per patient, we use Belief Propagation with sequential updates.

To learn the parameters of the observation factors we use the expectation-maximization (EM) algorithm.

### How to Make IPAs

After inference, we output an IPA for each variable that has an 'active' molecular type.

We compute a log-likelihood ratio using the quantities:

$$L(i,a) = \log \left( \frac{P(D,x_i=a|\Phi)}{P(D,x_i\neq a|\Phi)} \right) - \log \left( \frac{P(x_i=a|\Phi)}{P(x_i\neq a|\Phi)} \right)$$
$$= \log \left( \frac{P(D|x_i=a,\Phi)}{P(D|x_i\neq a,\Phi)} \right).$$

We then compute a single IPA for gene i based on the log-likelihood ratio as:

$$IPA(i) = \begin{cases} L(i,1) & L(i,1) > L(i,-1) \text{ and } L(i,1) > L(i,0) \\ -L(i,-1) & L(i,-1) > L(i,1) \text{ and } L(i,-1) > L(i,0) \\ 0 & \text{otherwise.} \end{cases}$$

# Aside: Factor Graphs

Consider a scenario where we have four students who get together in pairs to work on the homework for a class. For various reasons, only the following pairs meet: Alice and Bob; Bob and Charles; Charles and Debbie; and Debbie and Alice. (Alice and Charles just can't stand each other, and Bob and Debbie had a relationship that ended badly.) The study pairs are shown in figure 3.10a.

In this example, the professor accidentally misspoke in class, giving rise to a possible misconception among the students in the class. Each of the students in the class may subsequently have figured out the problem, perhaps by thinking about the issue or reading the textbook. In subsequent study pairs, he or she may transmit this newfound understanding to his or her study partners. We therefore have four binary random variables, representing whether the student has the misconception or not. We assume that for each  $X \in \{A, B, C, D\}$ ,  $x^1$  denotes the case where the student has the misconception, and  $x^0$  denotes the case where he or she does not.

Because Alice and Charles never speak to each other directly, we have that A and C are conditionally independent given B and D. Similarly, B and D are conditionally independent given A and C. Can we represent this distribution (with these independence properties) using a BN? One attempt is shown in figure 3.10b. Indeed, it encodes the independence assumption that  $(A \perp C \mid \{B,D\})$ . However, it also implies that B and D are independent given only A, but dependent given both A and C. Hence, it fails to provide a perfect map for our target distribution. A second attempt, shown in figure 3.10c, is equally unsuccessful. It also implies that  $(A \perp C \mid \{B,D\})$ , but it also implies that B and D are marginally independent. It is clear that all other candidate BN structures are also flawed, so that this distribution does not have a perfect map.

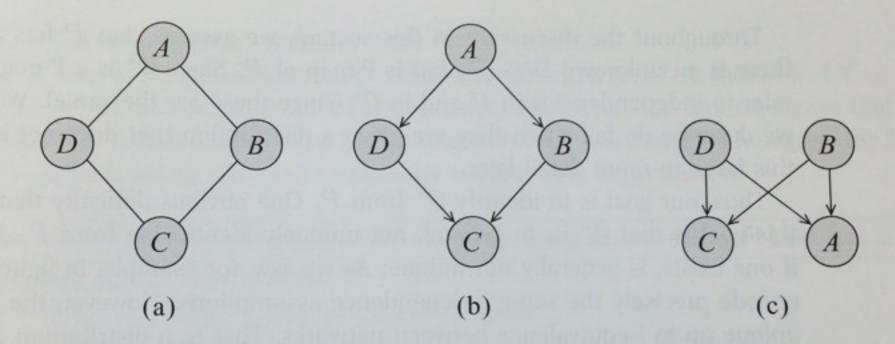


Figure 3.10 Attempted Bayesian network models for the Misconception example: (a) Study pairs over four students. (b) First attempt at a Bayesian network model. (c) Second attempt at a Bayesian network model.

# Draw Factor Graph

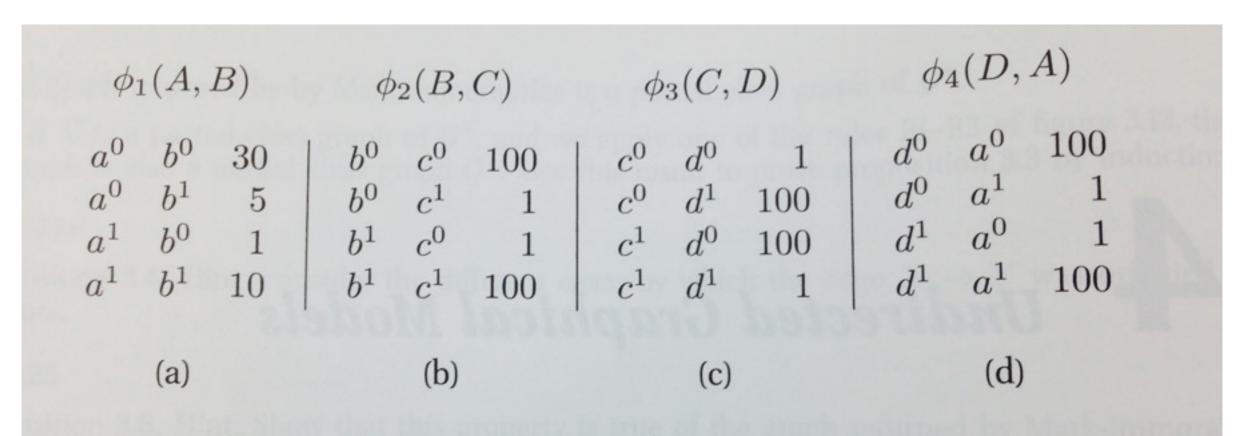


Figure 4.1 Factors for the Misconception example

	ssig	nme	nt	Unnormalized	Normalized		
$a^0$	$b^0$	$c^0$	$d^0$	300,000	0.04		
$a^0$	$b^0$	$c^0$	$d^1$	300,000	0.04		
$a^0$	$b^0$	$c^1$	$d^0$	300,000	0.04		
$a^0$	$b^0$	$c^1$	$d^1$	30	$4.1 \cdot 10^{-6}$		
$a^0$	$b^1$	$c^0$	$d^0$	500	$6.9 \cdot 10^{-5}$		
$a^0$	$b^1$	$c^0$	$d^1$	500	$6.9 \cdot 10^{-5}$		
$a^0$	$b^1$	$c^1$	$d^0$	5,000,000	0.69		
$a^0$	$b^1$	$c^1$	$d^1$	500	$6.9 \cdot 10^{-5}$		
$a^1$	$b^0$	$c^0$	$d^0$	100	$1.4 \cdot 10^{-5}$		
$a^1$	$b^0$	$c^0$	$d^1$	1,000,000	0.14		
$a^1$	$b^0$	$c^1$	$d^0$	100	$1.4 \cdot 10^{-5}$		
$a^1$	$b^0$	$c^1$	$d^1$	100	$1.4 \cdot 10^{-5}$		
$a^1$	$b^1$	$c^0$	$d^0$	10	$1.4 \cdot 10^{-6}$		
$a^1$	$b^1$	$c^0$	$d^1$	100,000	0.014		
$a^1$	$b^1$	$c^1$	$d^0$	100,000	0.014		
$a^1$	$b^1$	$c^1$	$d^1$	100,000	0.014		

Figure 4.2 Joint distribution for the Misconception example. The unnormalized measure and the normalized joint distribution over A, B, C, D, obtained from the parameterization of figure 4.1. The value of the partition function in this example is 7,201,840.

# Too Tired to Merge These Slides

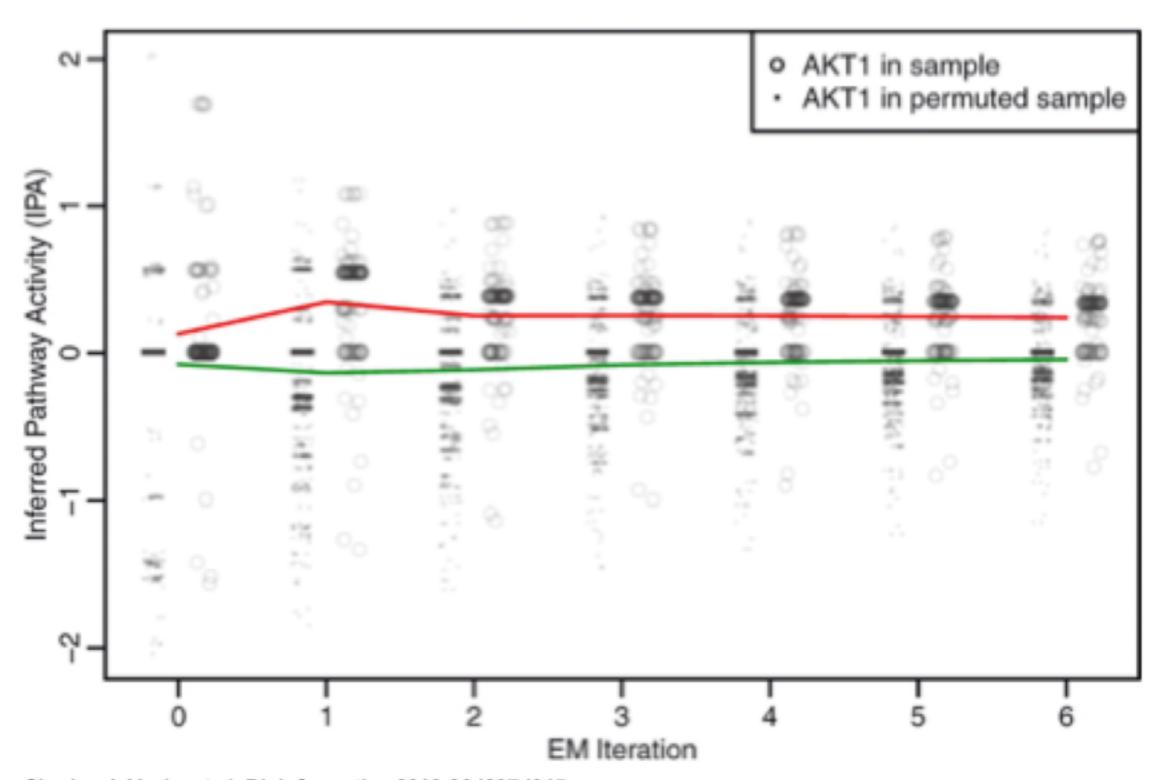
http://www.cedar.buffalo.edu/~srihari/CSE574/Chap8/ Ch8-GraphicalModelInference/Ch8.3.2-FactorGraphs.pdf

http://disi.unitn.it/~passerini/teaching/2010-2011/ MachineLearning/slides/09\_inference\_in\_bn/talk.pdf

http://www.cs.cmu.edu/~sandholm/cs15-780S11/slides/19-factor-graphs-mc.pdf

# Results

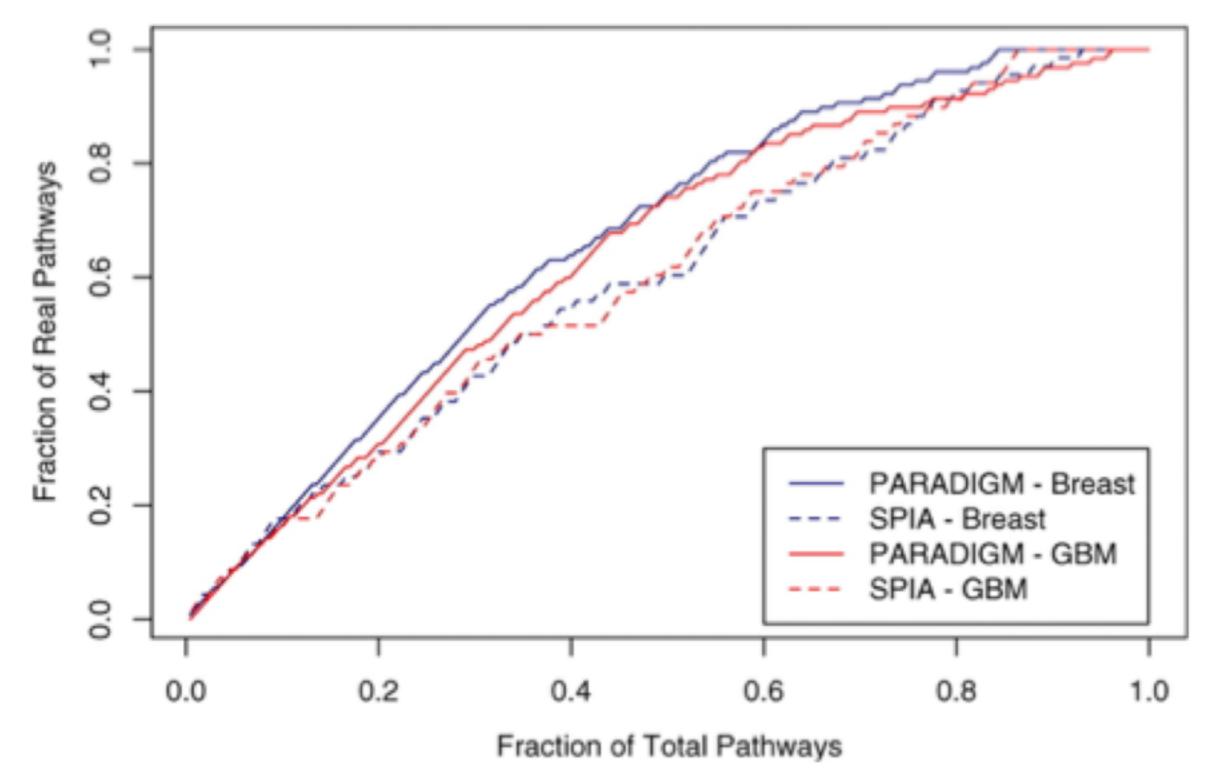
#### Learning parameters for AKT1.



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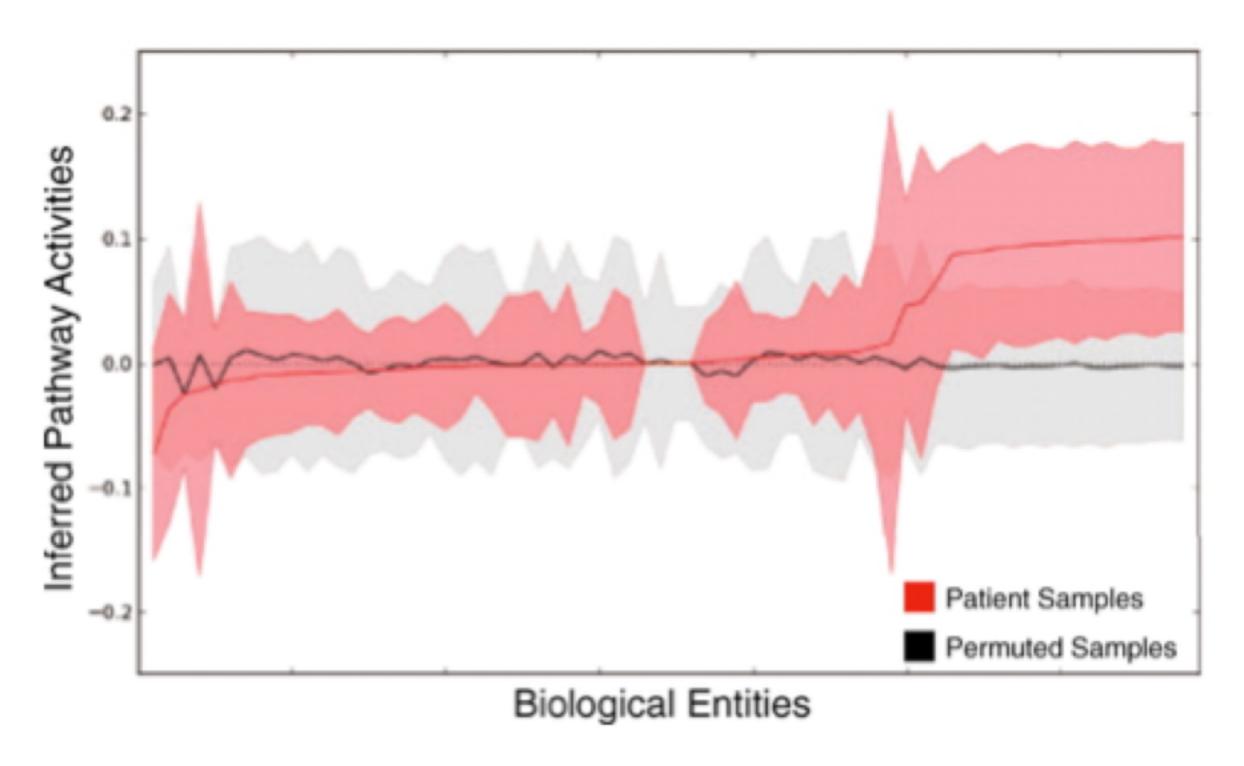
### Distinguishing decoy from real pathways with PARADIGM and SPIA. Decoy pathways were created by assigning a new gene name to each gene in a pathway.



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#### Patient sample IPAs compared with 'within' permutations for Class I PI3K signaling events mediated by Akt in breast cancer.



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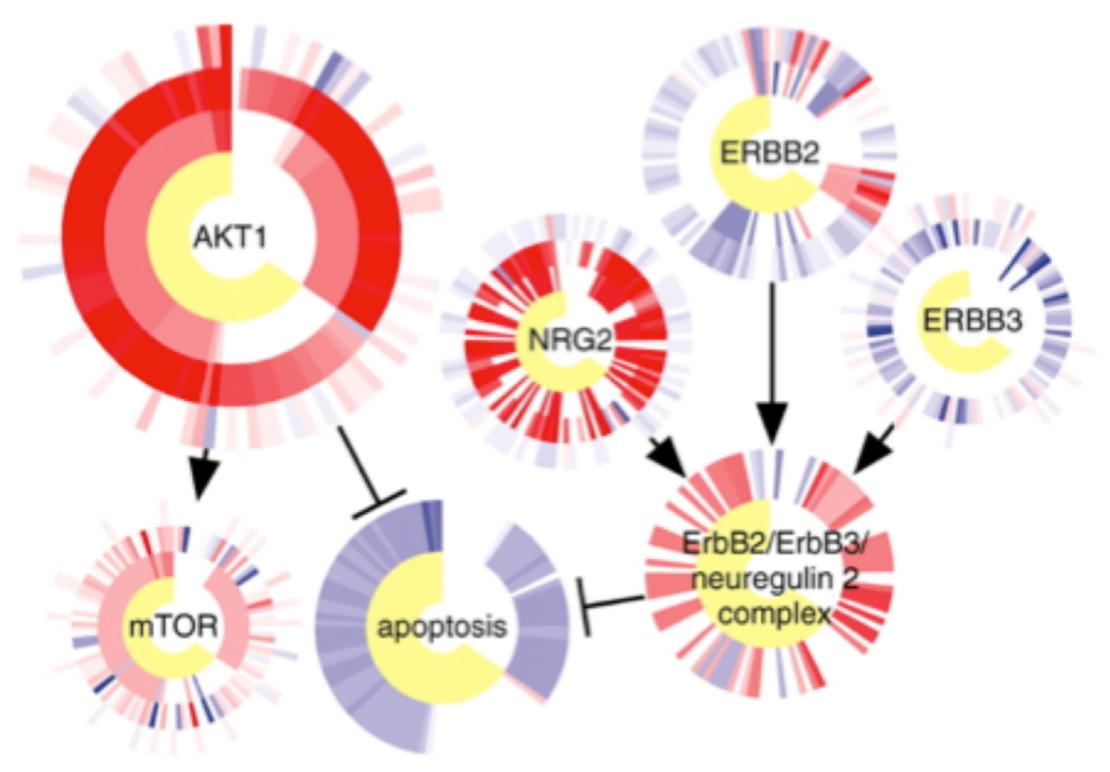


**Table 1.**Top PARADIGM pathways in breast cancer

**Table 2.**Top PARADIGM pathways in GBM

Rank	Name	Avg.a	SPIA?b	Rank	Name	Avg.a	SPIA?b
1	Class I PI3K signaling events mediated by Akt	20.7	No	1	Signaling by Ret tyrosine kinase	46.0	No
2	Nectin adhesion pathway	14.1	No	2	Signaling events activated by Hepatocyte GFR	43.7	No
3	Insulin-mediated glucose transport	13.8	No	3	Endothelins	42.5	Yes
4	ErbB2/ErbB3 signaling events	12.1	Yes	4	Arf6 downstream pathway	42.3	No
5	p75(NTR)-mediated signaling	11.5	No	5	Signaling events mediated by HDAC Class III	36.3	No
6	HIF-1-alpha transcription factor network	10.7	No	6	FOXM1 transcription factor network	35.9	Yes
7	Signaling events mediated by PTP1B	10.7	No	7	IL6-mediated signaling events	33.2	No
8	Plasma membrane estrogen receptor signaling	10.6	Yes	8	FoxO family signaling	31.3	No
9	TCR signaling in naive CD8+ T cells	10.6	No	9	LPA receptor mediated events	30.7	Yes
10	Angiopoietin receptor Tie2-mediated signaling	10.1	No	10	ErbB2/ErbB3 signaling events	30.1	No
11	Class IB PI3K non-lipid kinase events	10.0	No	11	Signaling mediated by p38-alpha and p38-beta	28.1	No
13	Osteopontin-mediated events	9.9	Yes	12	HIF-1-alpha transcription factor network	27.6	Yes
12	IL4-mediated signaling events	9.8	No	13	Non-genotropic Androgen signaling	27.3	No
14	Endothelins	9.8	No	14	p38 MAPK signaling pathway	27.2	No
15	Neurotrophic factor-mediated Trk signaling	9.7	No	15	IL2 signaling events mediated by PI3K	26.9	No

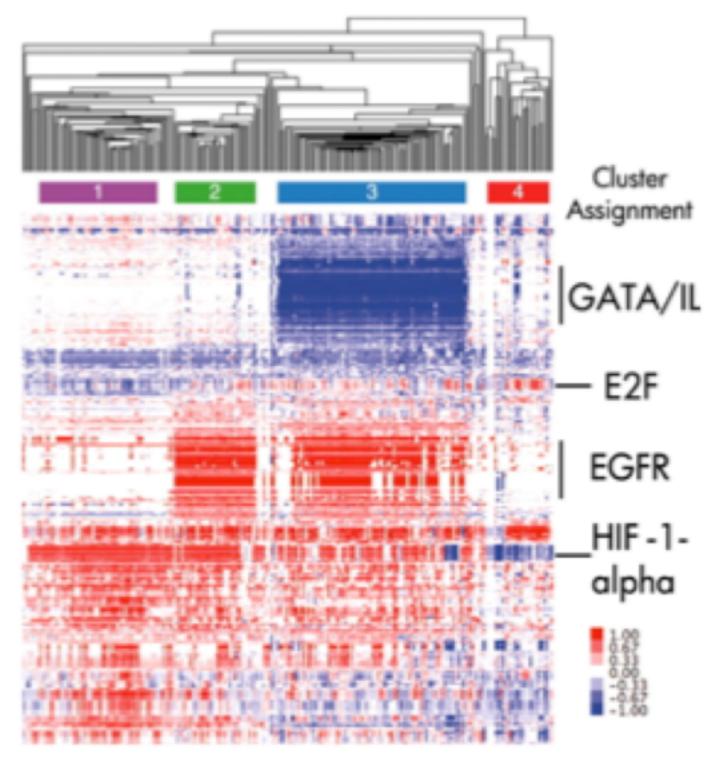
#### CircleMap display of the ErbB2 pathway.



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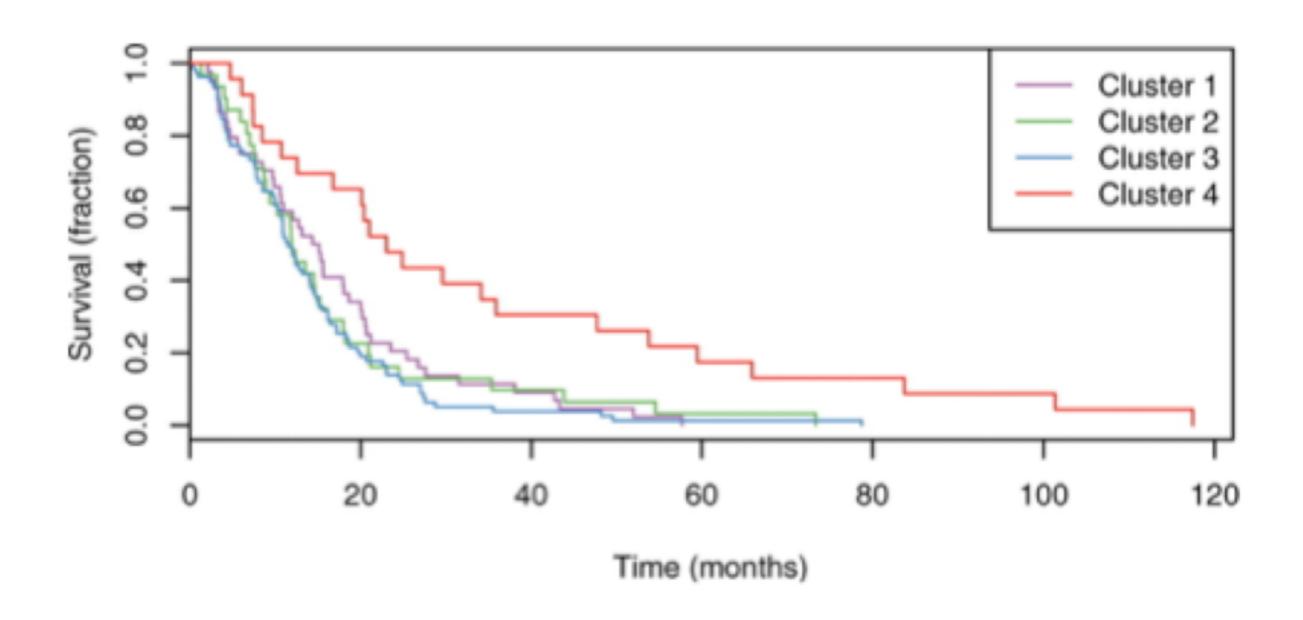
### Clustering of IPAs for TCGA GBM. Each column corresponds to a single sample, and each row to a biomolecular entity.



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#### Kaplan-Meier survival plots for the clusters from Figure 8.



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## Future Work