

# QTL mapping 2: Special topics

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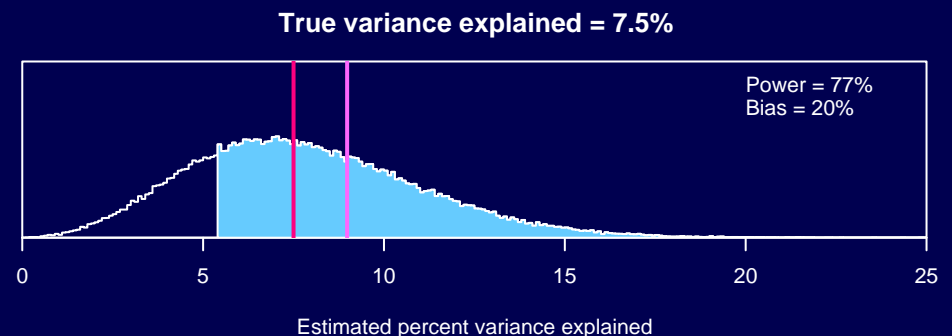
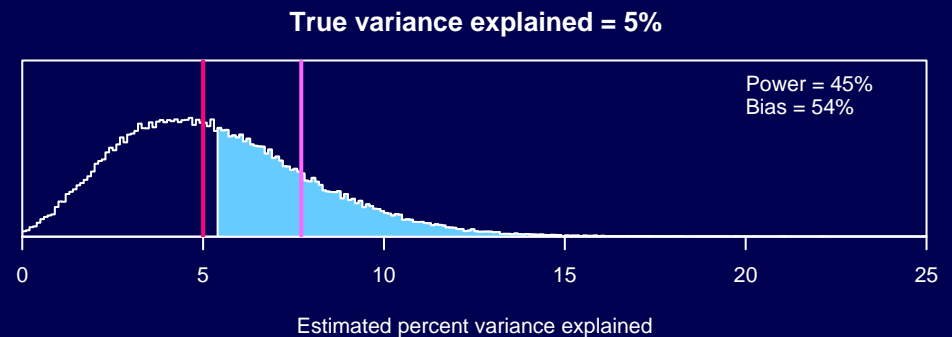
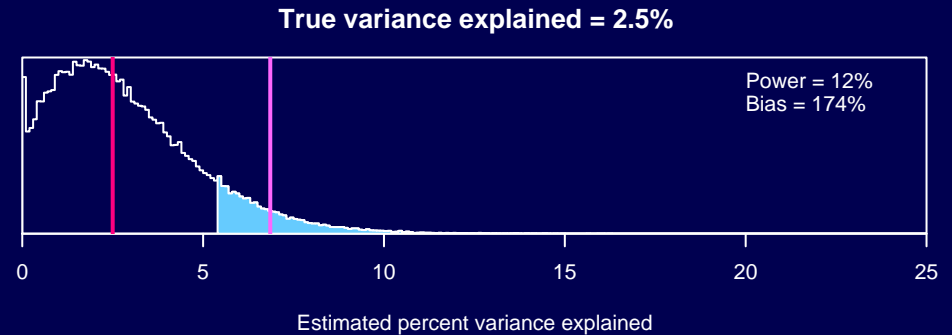
[kbroman.org](http://kbroman.org)

[github.com/kbroman](https://github.com/kbroman)

@kwbroman

# Selection bias

- The estimated effect of a QTL will vary somewhat from its true effect.
- Only when the estimated effect is large will the QTL be detected.
- Among those experiments in which the QTL is detected, the estimated QTL effect will be, on average, larger than its true effect.
- This is **selection bias**.
- Selection bias is largest in QTLs with small or moderate effects.
- The true effects of QTLs that we identify are likely smaller than was observed.



# Implications

- Estimated % variance explained by identified QTLs
- Repeating an experiment
- Congenics (aka near isogenic lines)
- Marker-assisted selection

# Non-normal traits

- Standard interval mapping assumes normally distributed residual variation. (Thus the phenotype distribution is a mixture of normals.)
- **In reality**: we see dichotomous traits, counts, skewed distributions, outliers, and all sorts of odd things.
- Interval mapping, with LOD thresholds derived from permutation tests, generally performs just fine anyway.
- Alternatives to consider:
  - Nonparametric approaches (Kruglyak & Lander 1995)
  - Transformations (*e.g.*, log, square root, normal quantiles)
  - Specially-tailored models (*e.g.*, a generalized linear model, the Cox proportional hazard model, and the two-part model in Broman 2003)

# Haley-Knott regression

A quick approximation to Interval Mapping.

$$E(y_i|q_i) = \mu_q$$

$$\begin{aligned} E(y_i|M_i) &= E[ E(y_i|q_i) |M_i] = \sum_j \Pr(q = j|M_i)\mu_j \\ &= \sum_j p_{ij}\mu_j \end{aligned}$$

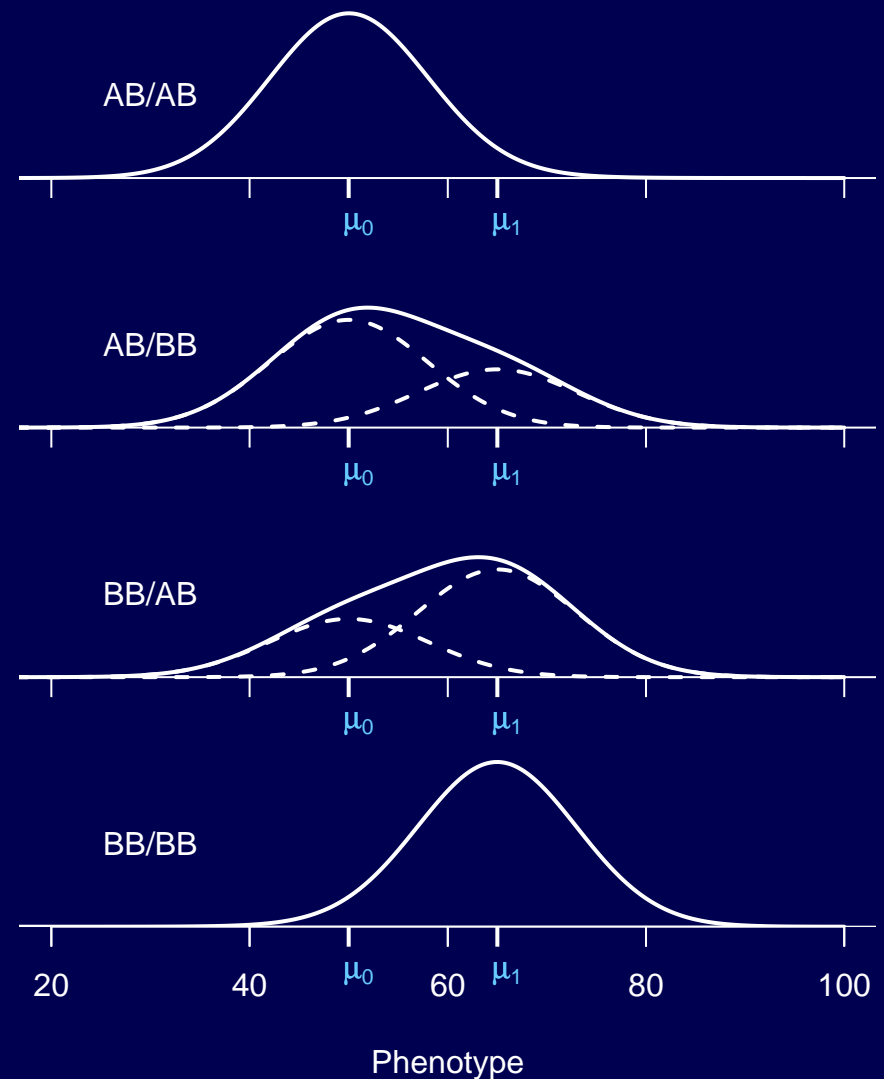
Regress  $y$  on  $p_i$ , pretending the residual variation is normally distributed (with constant variance).

$$\text{LOD} = \frac{n}{2} \log_{10} \left( \frac{\text{RSS}_0}{\text{RSS}_1} \right)$$

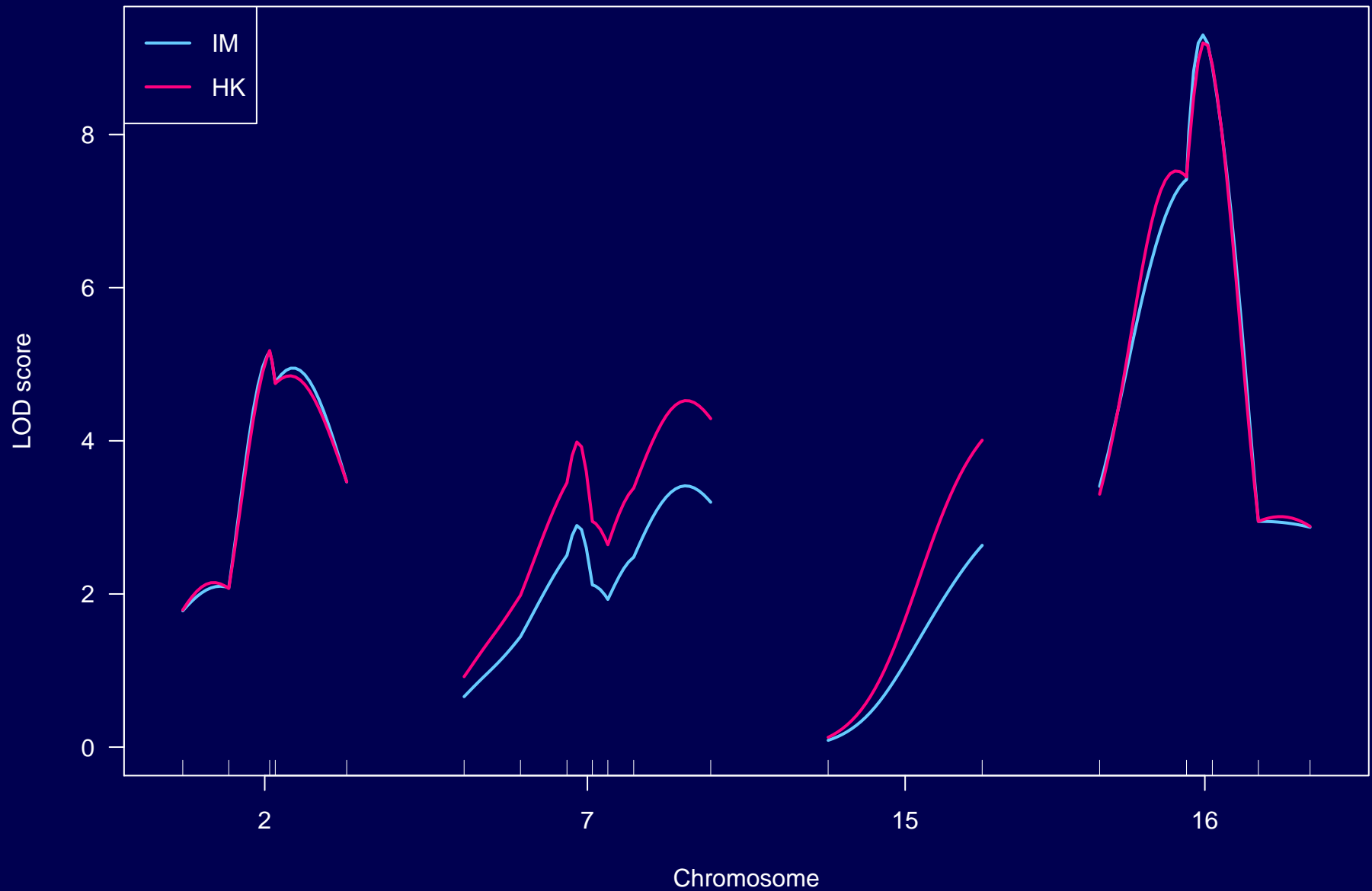
# The normal mixtures



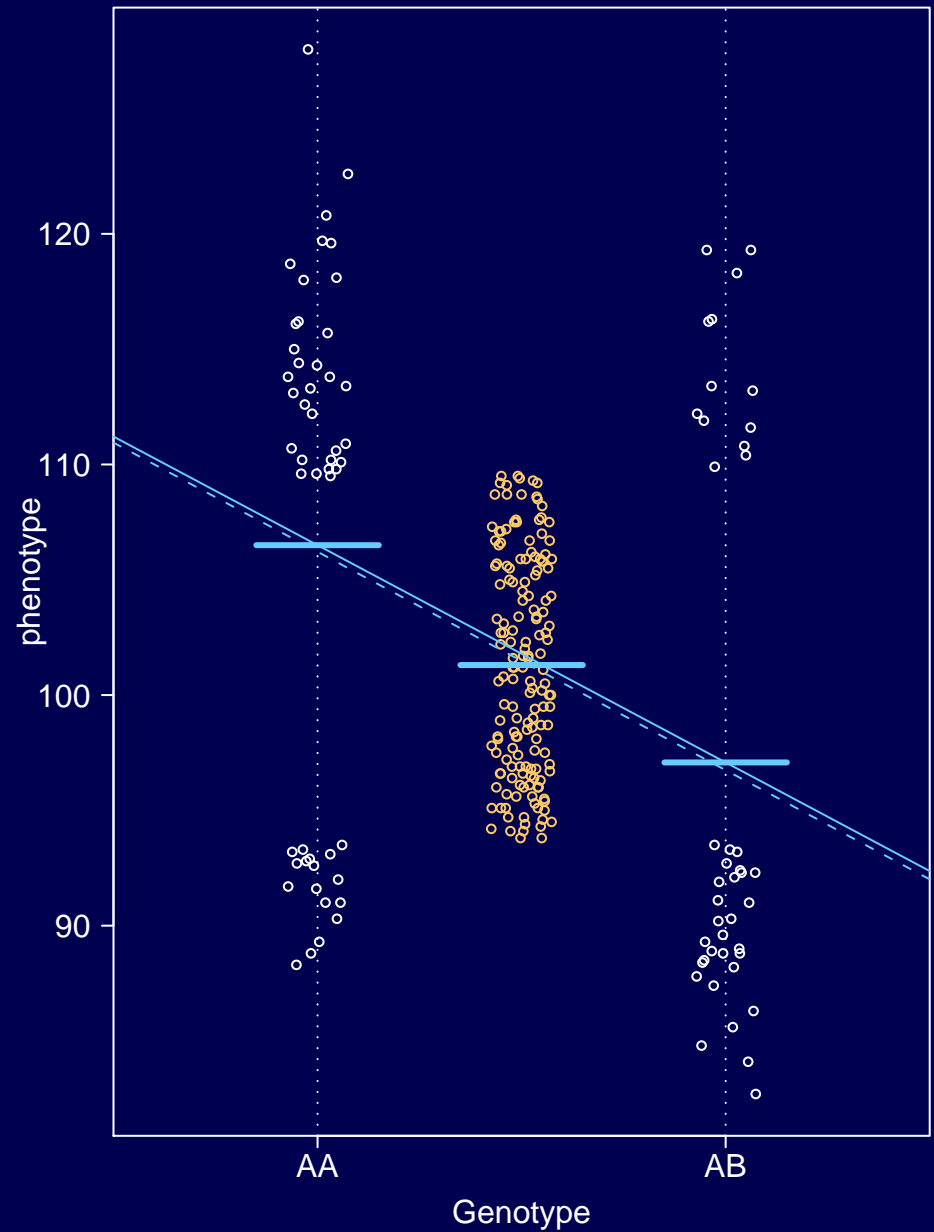
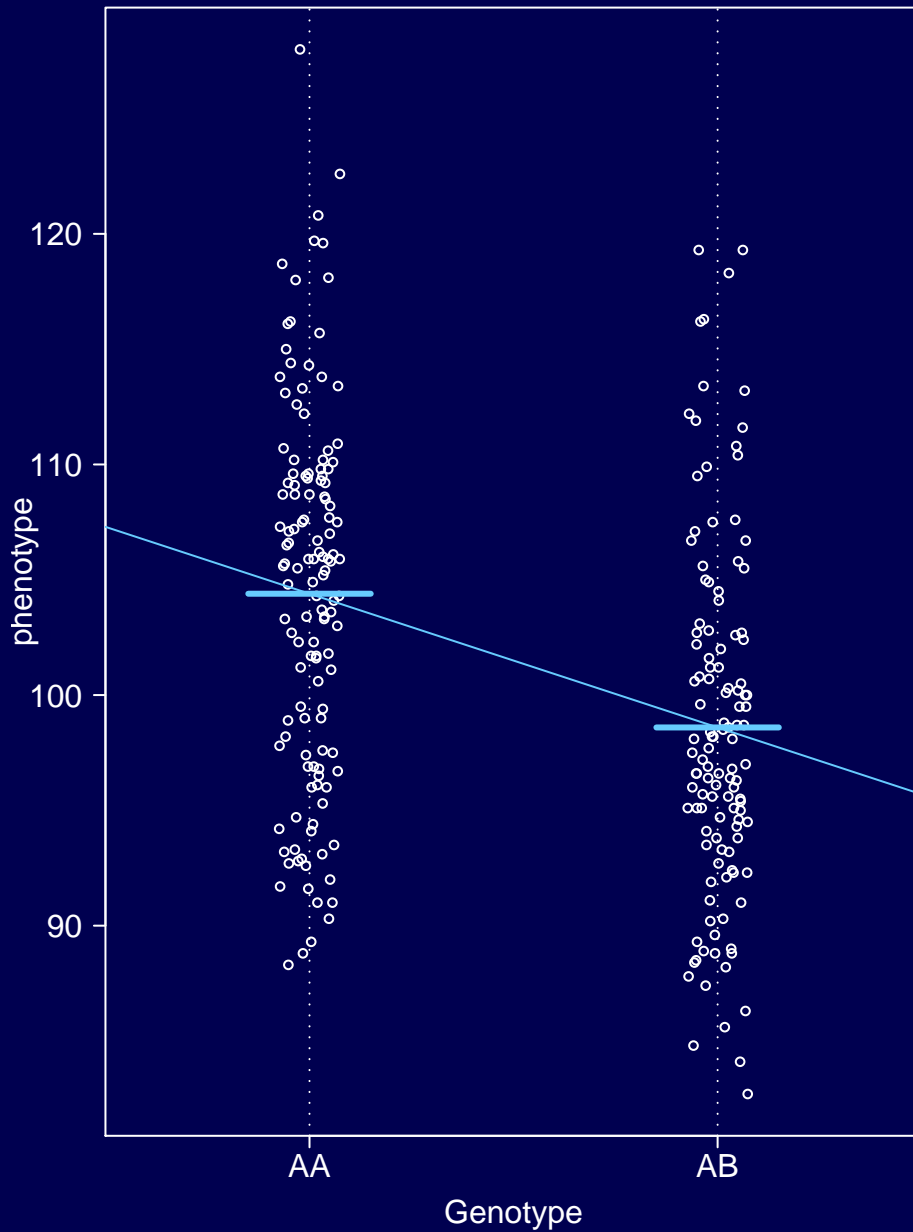
- Two markers separated by 20 cM, with the QTL closer to the left marker.
- The figure at right shows the distributions of the phenotype conditional on the genotypes at the two markers.
- The dashed curves correspond to the components of the mixtures.



# Haley-Knott results



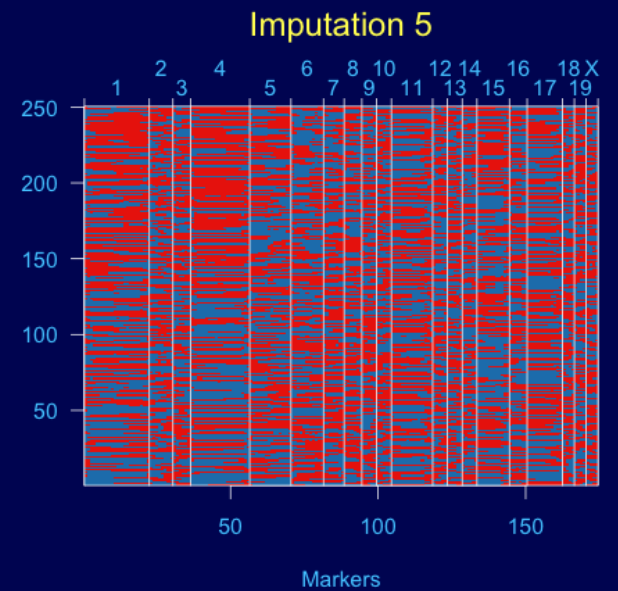
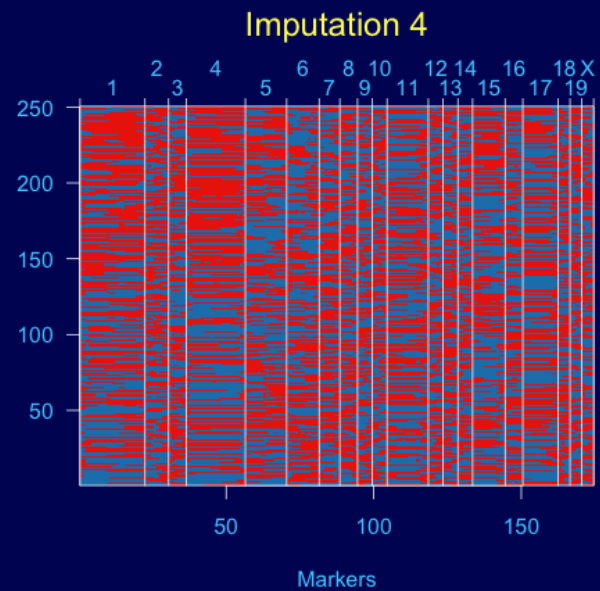
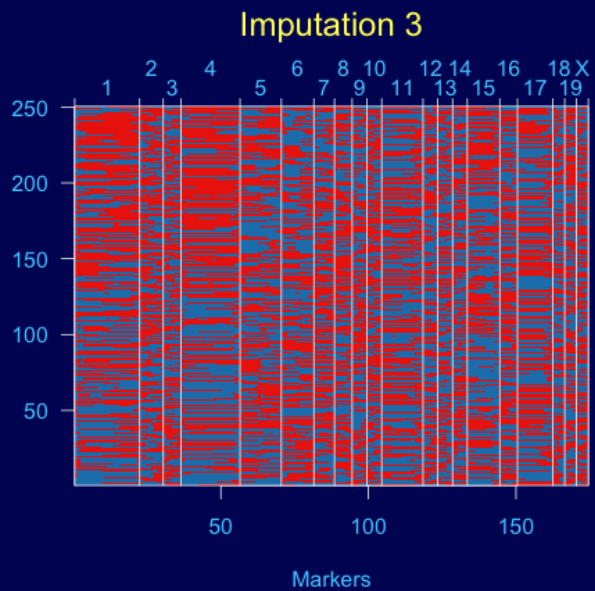
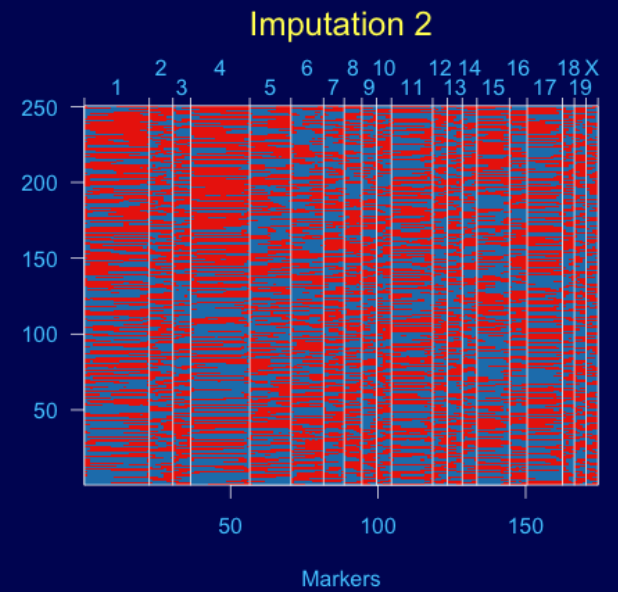
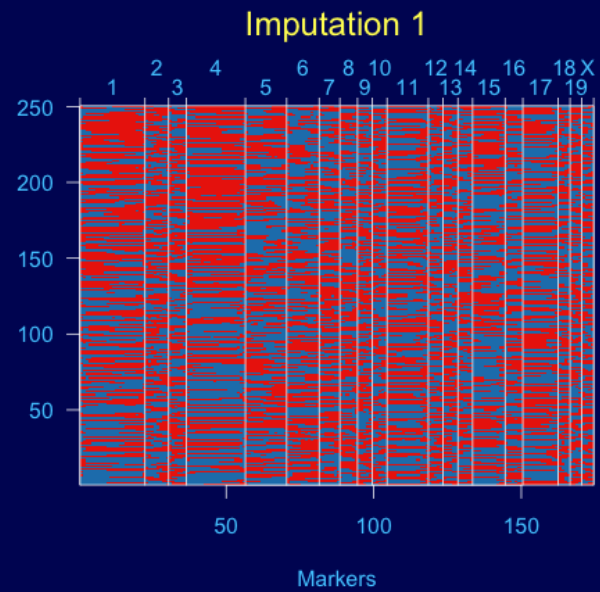
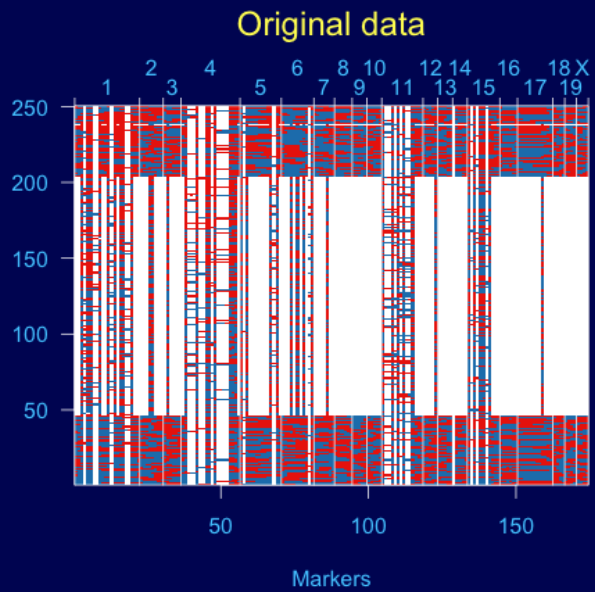
# H-K with selective genotyping



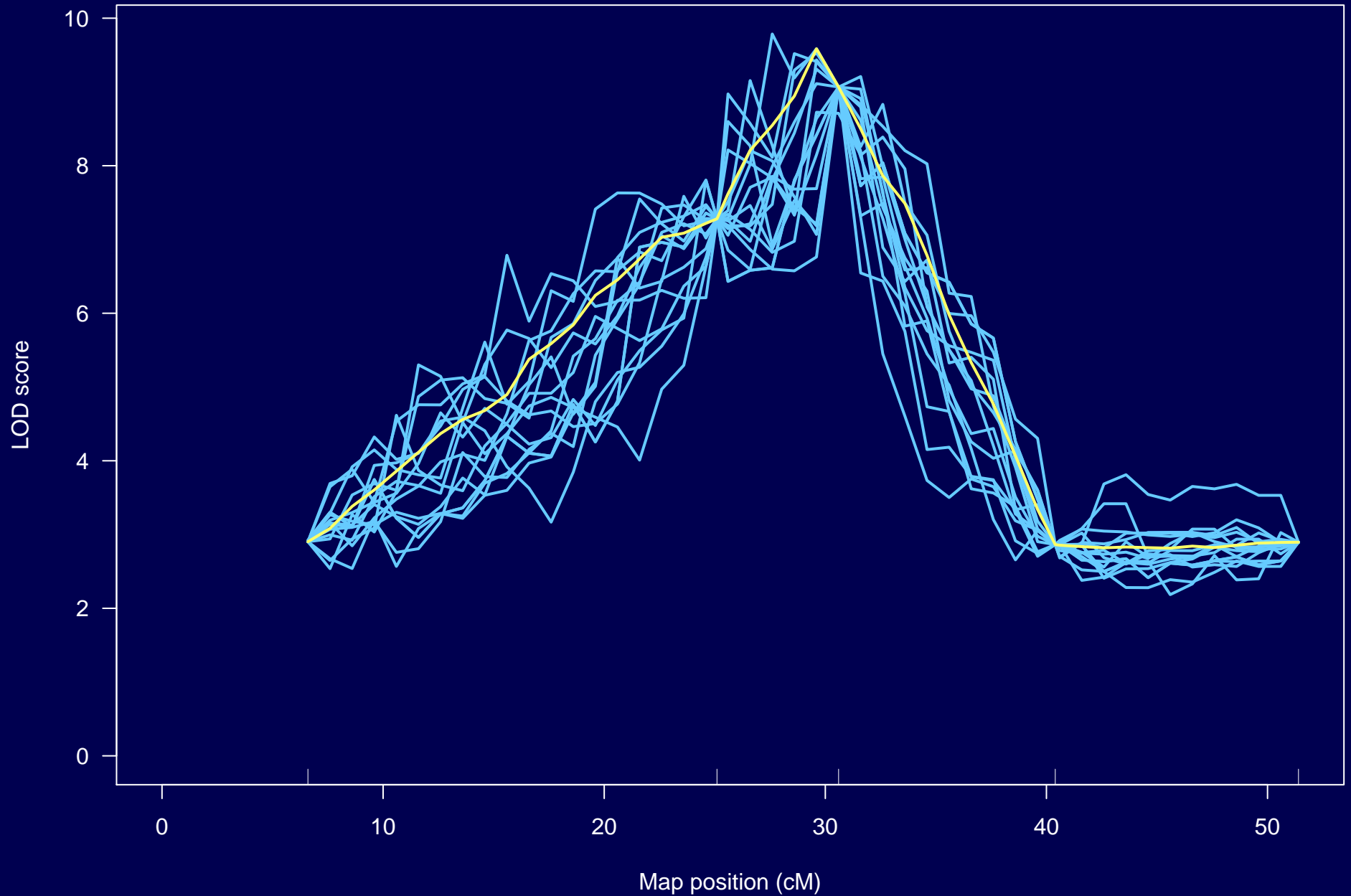




# Multiple imputations



# Imputation LOD curves



# Summary comparison

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Approach	Speed	Extensibility	Stability	Missing data	Parallelization
HK	++	+	+	-	++
EM	+	-	-	+	-
Imputation	-	+	+	+	+

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# Covariates

- **Examples:** treatment, sex, age, weight
- Control residual variation → increase power
- Look for QTL × covariate interactions

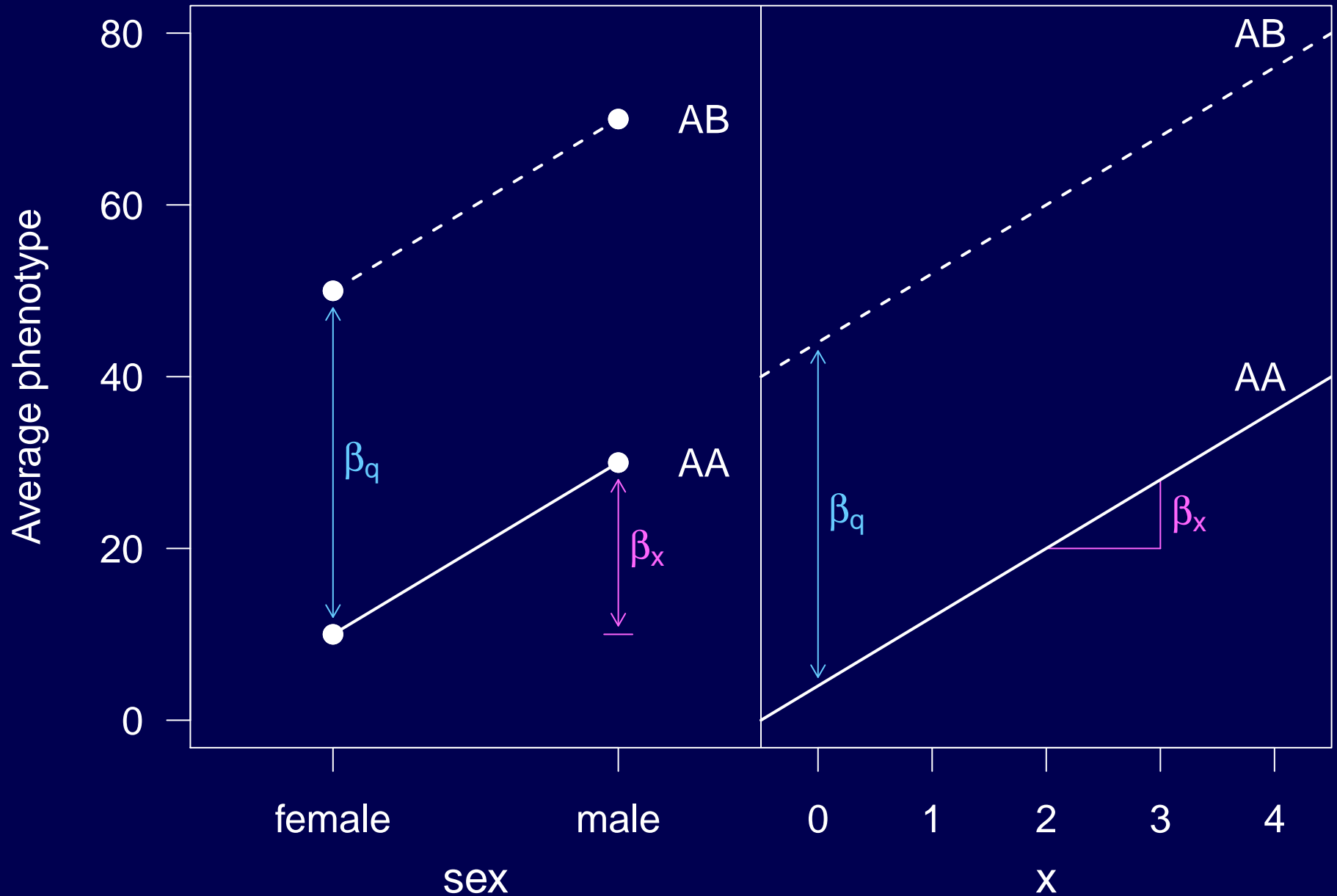
# Additive covariate

$$H_0 : y = \mu + \beta_x x + \epsilon$$

$$H_a : y = \mu + \beta_x x + \beta_q q + \epsilon$$

- If covariate has strong effect on the phenotype, accounting for it can give improved power to detect QTL.
- In permutations, keep phenotype and covariate together
- Use care when the covariate is another phenotype

# Additive covariate



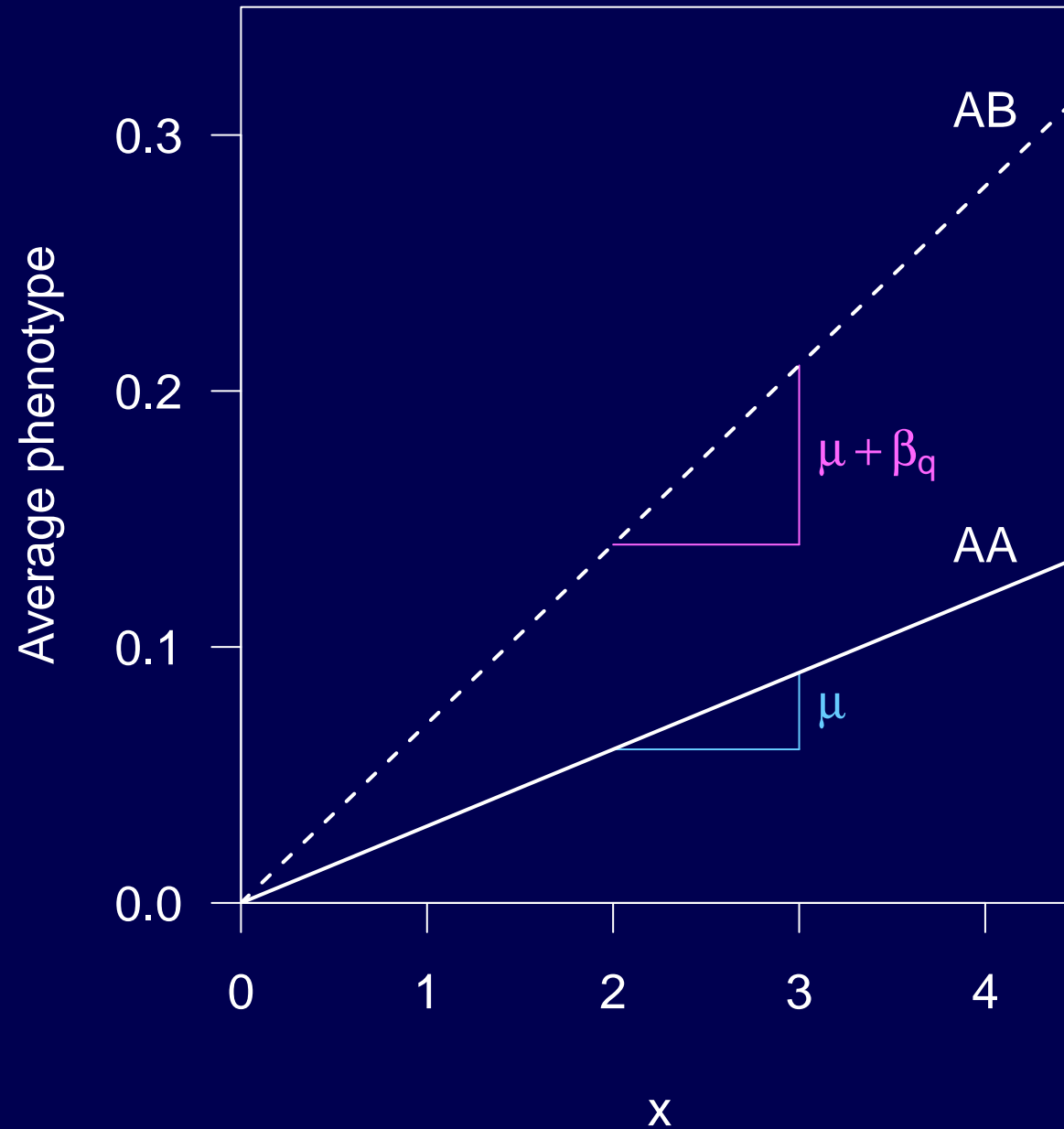
# Adjust then scan?

- Consider adjusted phenotype  $y' = y/x$
- The QTL model is  $(y/x) = \mu + \beta_q q + \epsilon$
- Equivalently

$$y = \begin{cases} \mu x + \epsilon' & \text{if } q = 0 \\ (\mu + \beta_q)x + \epsilon' & \text{if } q = 1 \end{cases}$$



# Adjust then scan?



# Interactive covariate

$$H_0 : y = \mu + \beta_x x + \epsilon$$

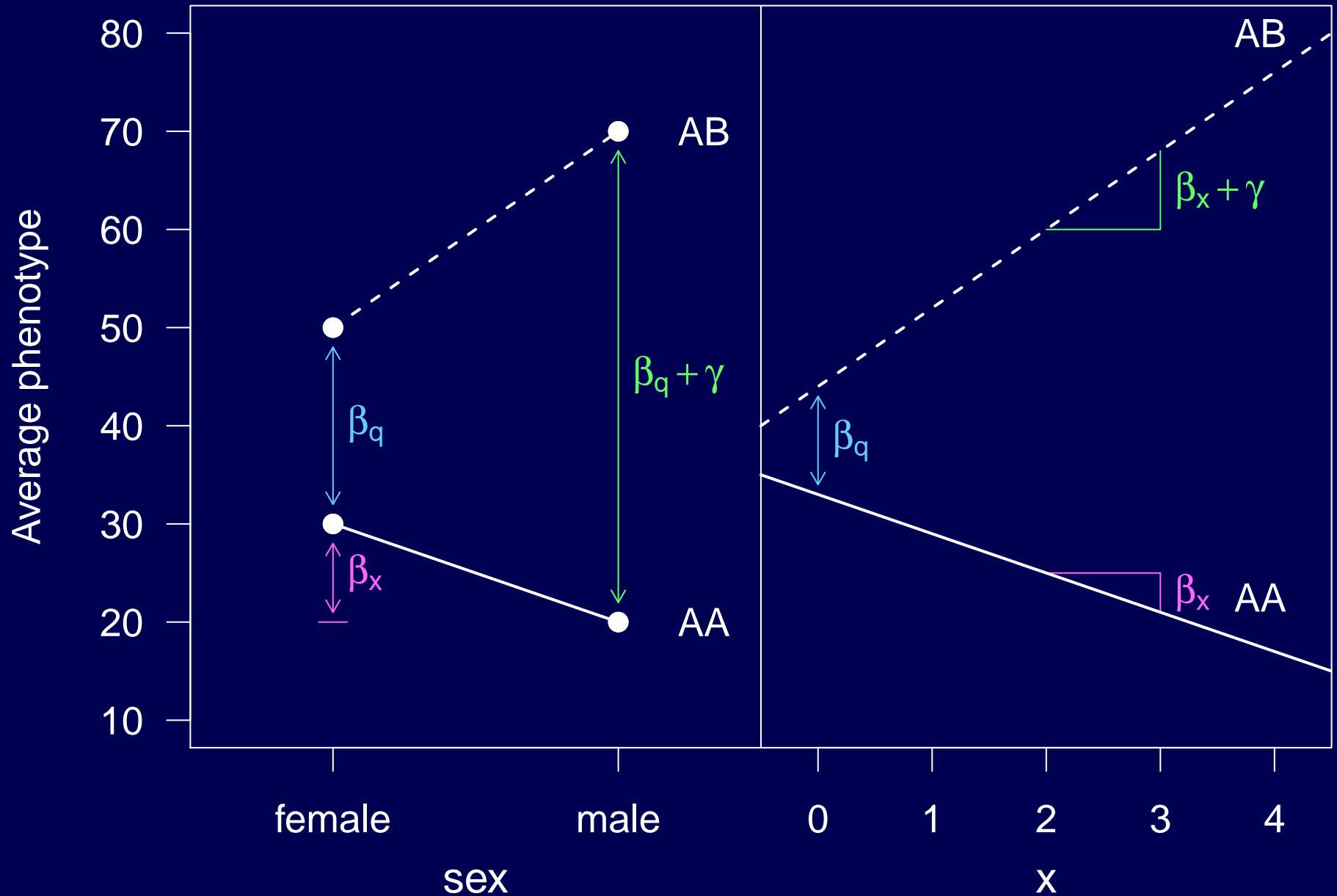
$$H_a : y = \mu + \beta_x x + \beta_q q + \epsilon$$

$$H_i : y = \mu + \beta_x x + \beta_q q + \gamma xq + \epsilon$$

Can consider 3 LOD scores:

- $LOD_a$  comparing  $H_a$  and  $H_0$
- $LOD_f$  comparing  $H_i$  and  $H_0$
- $LOD_i$  comparing  $H_i$  and  $H_a$

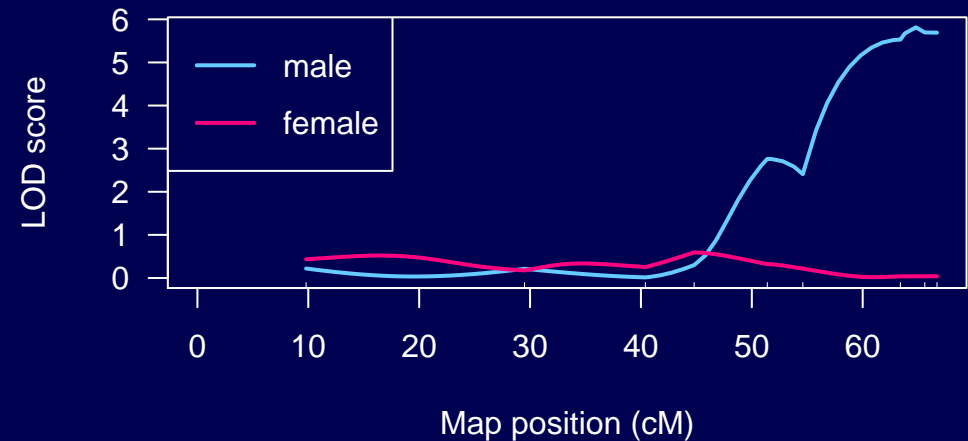
# Interactive covariate



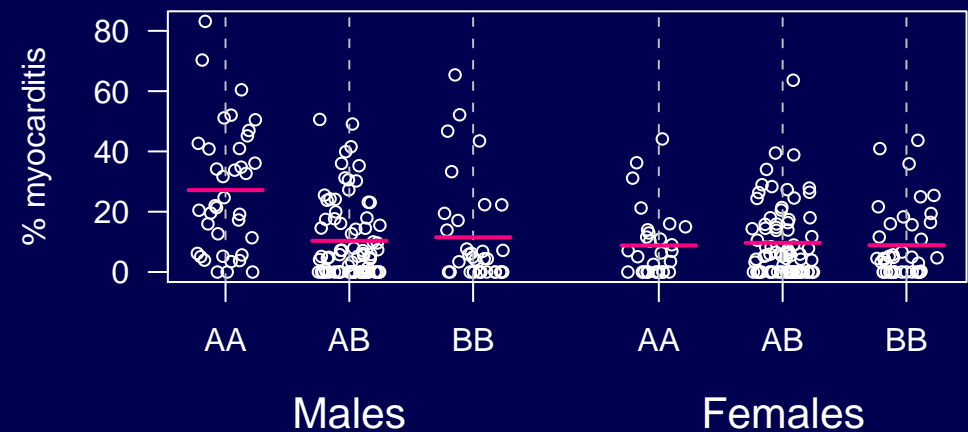
# Split on sex?

- Informative, understandable
- But tempting to falsely conclude “**sex-specific QTL**”
- Absence of evidence **is not evidence of absence**.
- Use explicit test of QTL  $\times$  sex interaction

Chromosome 6

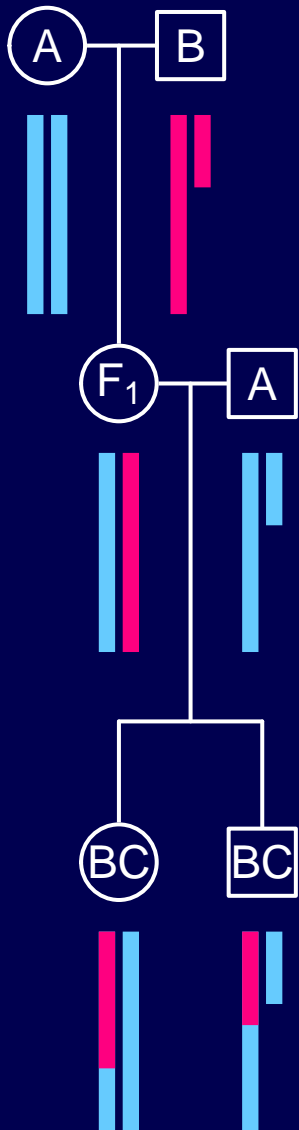


D6Mit373

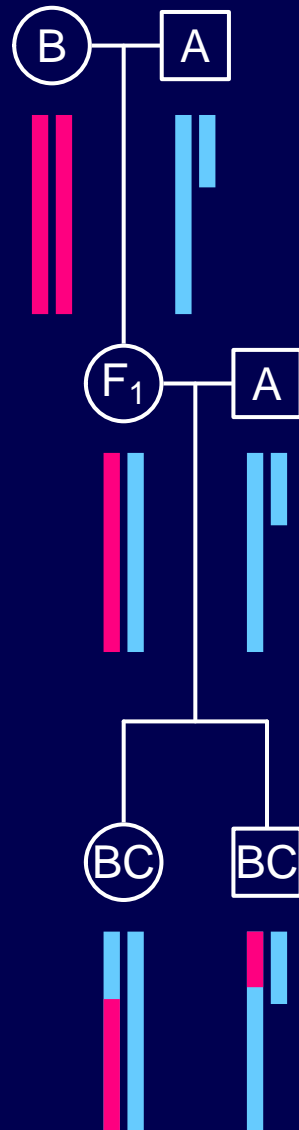


# X chr in backcross

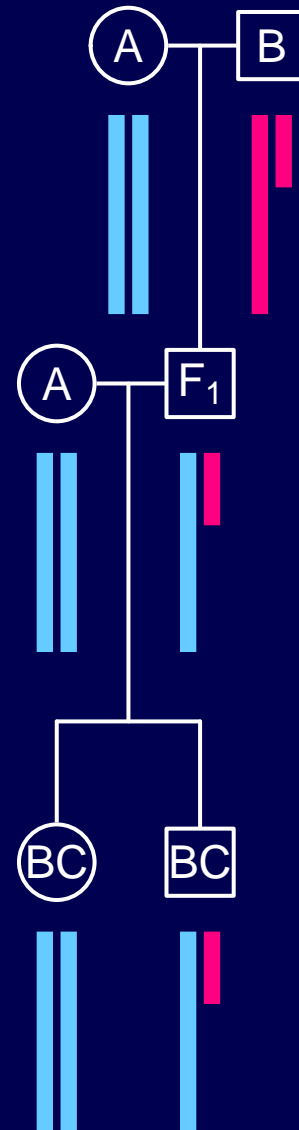
$(A \times B) \times A$



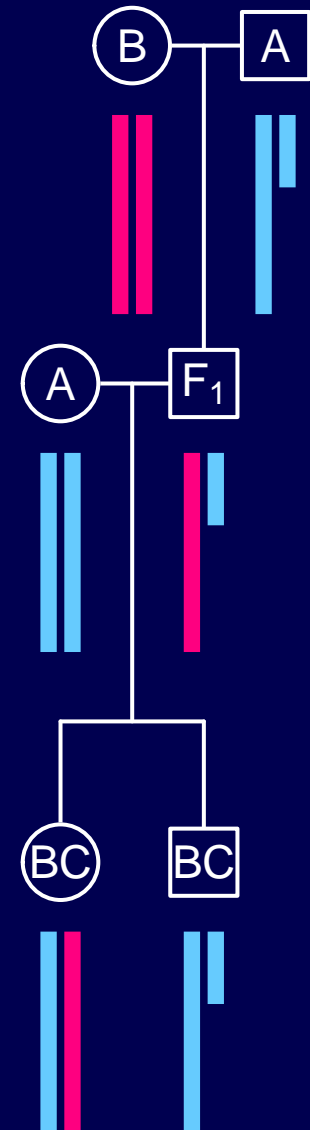
$(B \times A) \times A$



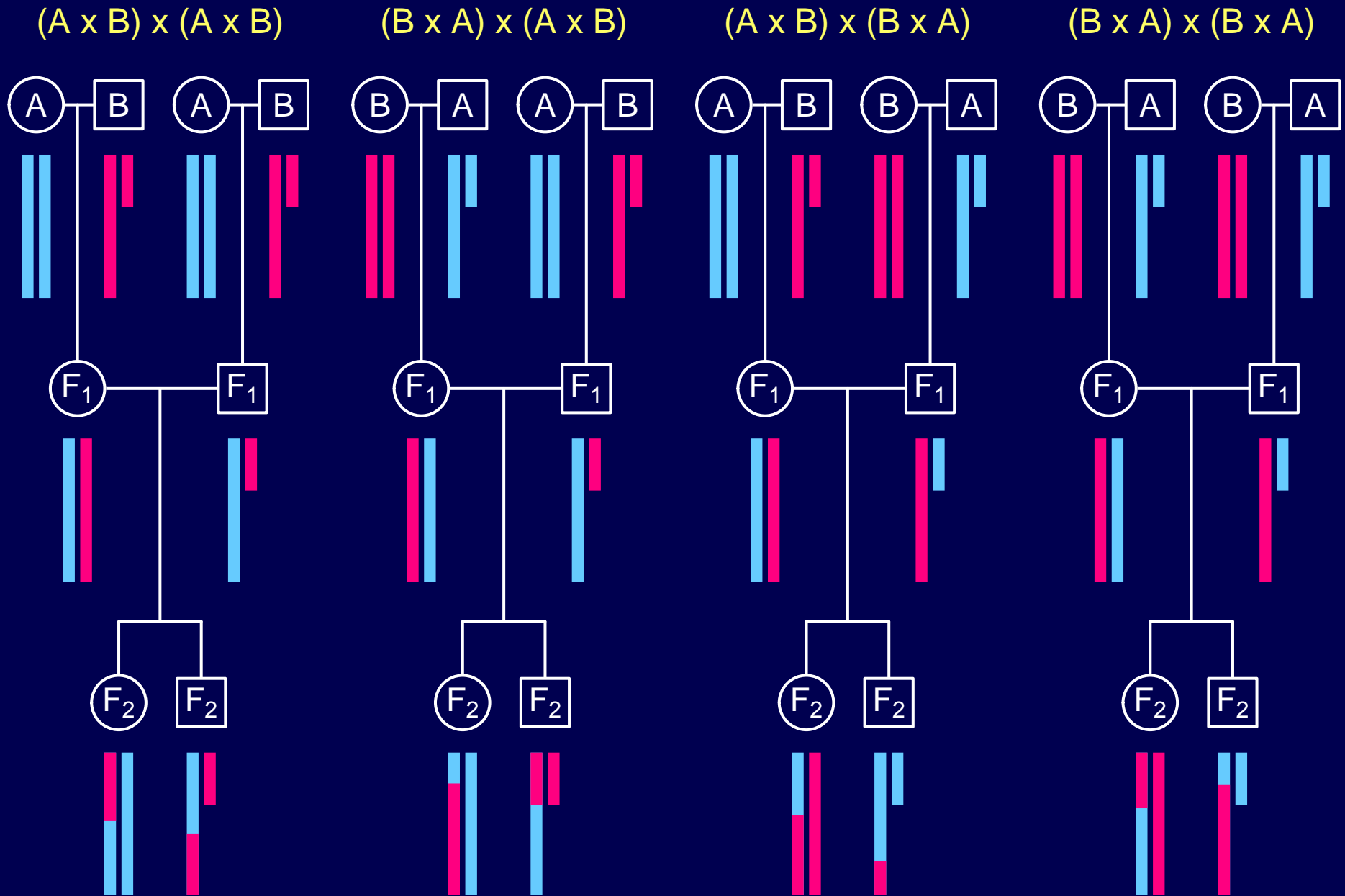
$A \times (A \times B)$



$A \times (B \times A)$



# X chr in intercross



# Example

Intercross: both dir, both sexes

♀ forward	AA or AB	
♀ reverse		AB or BB
♂ forward		AY or BY
♂ reverse		AY or BY

# Principles

- Sex- or cross-direction-difference in the phenotype shouldn't lead to spurious linkage on the X chromosome
- Simple as possible
- Null nested within alternative



# Approach

Cross	Direction	Sexes	Contrasts	H <sub>0</sub>	df
BC		both	AA:AB:AY:BY	♀:♂	2
BC		♀	AA:AB	grand mean	1
BC		♂	AY:BY	grand mean	1
F <sub>2</sub>	both	both	AA:AB <sub>f</sub> :AB <sub>r</sub> :BB:AY:BY	♀ <sub>f</sub> :♀ <sub>r</sub> :♂	3
F <sub>2</sub>	both	♀	AA:AB <sub>f</sub> :AB <sub>r</sub> :BB	♀ <sub>f</sub> :♀ <sub>r</sub>	2
F <sub>2</sub>	both	♂	AY:BY	grand mean	1
F <sub>2</sub>	one	both	AA:AB:AY:BY	♀:♂	2
F <sub>2</sub>	one	♀	AA:AB	grand mean	1
F <sub>2</sub>	one	♂	AY:BY	grand mean	1

# Chromosome-specific thresholds

Let  $\alpha_i$  = false positive rate for chromosome  $i$ .

We need  $1 - \alpha = \prod(1 - \alpha_i)$

For example,  $\alpha_1 = \alpha$  and  $\alpha_j = 0$  for  $j \neq 1$

The usual method: constant LOD threshold  
(i.e., constant power)

My approach:  $\alpha_i \propto L_i$  where  $L_i$  = length of chr  $i$

Similar and more convenient:  $\alpha_i = (1 - \alpha)^{L_i/L}$

# Data diagnostics

- Plot phenotypes
- Look for sample duplicates
- Look for excessive missing data
- Investigate segregation distortion
- Verify genetic maps/marker positions
- Look for genotyping errors
- Look at counts of crossovers

# References

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