

QTL Mapping II:

Model Selection

Karl W Broman

Department of Biostatistics
Johns Hopkins University

`kbroman@jhsph.edu`

`www.biostat.jhsph.edu/~kbroman`

Outline

- The model selection problem:
 - Class of models
 - Compare models
 - Search model space
 - Assess the performance of a procedure
- A simulation study

Hypothesis testing?

- In the past, QTL mapping has been regarded as a task of **hypothesis testing**.

Is this a QTL?

Much of the focus has been on adjusting for test multiplicity.

- It is better to view the problem as one of **model selection**.

What set of QTLs are well-supported?

Is there evidence for QTL-QTL interactions?

Model = a defined set of QTLs and QTL-QTL interactions (and possibly covariates and QTL-covariate interactions).

Perfect data situation

To ease discussion, we'll focus on a simple special case:

- Complete marker genotype data
- Markers are only putative QTLs
- Normally distributed residuals

Example model (in a backcross):

$$y_i = \mu + \Delta_1 q_{i1} + \Delta_2 q_{i2} + \Delta_3 q_{i3} + \epsilon_i \quad \epsilon_i \text{ are iid } N(0, \sigma^2)$$

q_j are 0/1 variables (QTL genotypes)

μ, Δ 's are parameters, estimated by least squares

Fitted values: $\hat{y}_i = \hat{\mu} + \hat{\Delta}_1 q_{i1} + \hat{\Delta}_2 q_{i2} + \hat{\Delta}_3 q_{i3}$

RSS = $\sum_i (y_i - \hat{y}_i)^2$ indicates model fit.

Model selection

1. Class of models
2. Compare models
3. Search model space
4. Assess performance of a procedure

Note:

2 and 4 are much the same.

There might be a 0th item: [Method for model fitting](#).
(e.g., imputation, EM, etc.)

Class of models

- Additive models

$$y = \mu + \sum_j \Delta_j q_j + \epsilon$$

- Also pairwise interactions

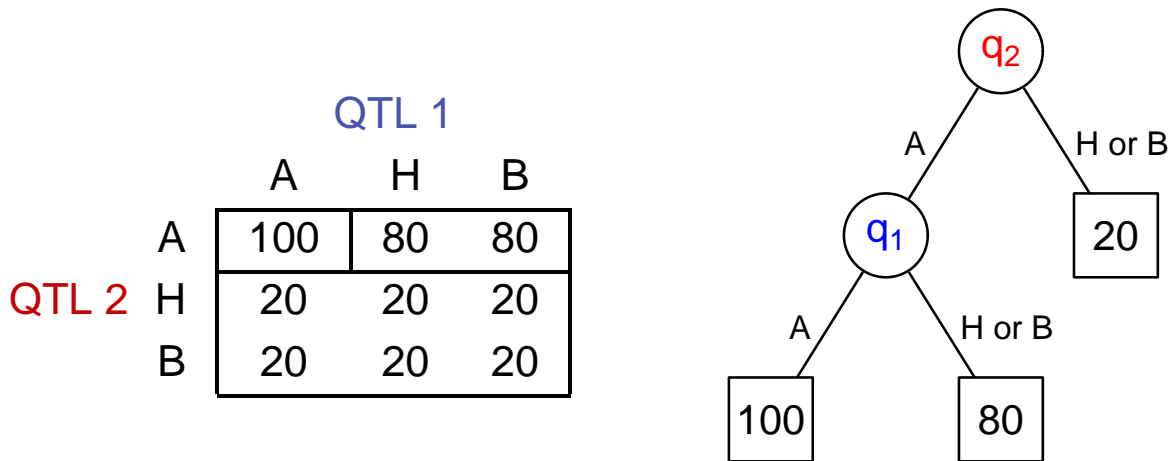
Preserve hierarchy: if include interaction, also include both main effects.

- Also higher-order interactions

Again, preserve hierarchy.

- Regression trees

Example regression tree



Intercrosses: class of models

- Always bring in both degrees of freedom with a QTL

or

Try to distinguish additivity/dominance/recessiveness?

A	H	B
---	---	---

- Always bring in all four d.f. with a QTL:QTL interaction

or

Try to distinguish ways to subdivide the 3×3 table?

	A	H	B
A			
H			
B			

Compare models

- Imagine you could fit all possible models; which would you like best?
- This issue is like LOD thresholds, but more complicated.
- For models with the same number of parameters (QTLs and interactions), we prefer that with the “best fit” (smallest RSS or largest likelihood).
- If you fit more parameters, you’ll get a “better fit.”
 - How much better, before including additional terms?
 - I like a form of penalized likelihood.
- Note: Bayesians must also confront this issue (through the prior on models).

The additive QTL case

n backcross mice; M markers

x_{ij} = genotype (1/0) of mouse i at marker j

y_i = phenotype (trait value) of mouse i

$$y_i = \mu + \sum_{j=1}^M \Delta_j x_{ij} + \epsilon_i \quad \text{Which } \Delta_j \neq 0?$$

$$\text{BIC}_\delta = \log \text{RSS} + \text{no. markers} \times \left(\delta \times \frac{\log n}{n} \right)$$

Why BIC_δ ?

- For a fixed no. markers, letting $n \rightarrow \infty$, BIC_δ is **consistent**.
- There exists a prior (on models + coefficients) for which BIC_δ is the **$-\log$ posterior**.
- BIC_δ is essentially equivalent to use of a threshold on the conditional LOD score
- It performs well.

$BIC_\delta \longleftrightarrow$ conditional LOD

Conditional LOD score:

$$\text{LOD}(x_k^* | x_1^*, \dots, x_{k-1}^*) = \frac{n}{2} \log_{10} \left\{ \frac{\text{RSS}(x_1^*, \dots, x_{k-1}^*)}{\text{RSS}(x_1^*, \dots, x_k^*)} \right\}$$

Minimizing BIC_δ is approximately equivalent to choosing the largest **k** such that

$$\text{LOD}(x_k^* | x_1^*, \dots, x_{k-1}^*) \geq \frac{\delta}{2} \log_{10} n$$

Choice of δ

Smaller δ : include more loci; higher false positive rate

Larger δ : include fewer loci; lower false positive rate

Let $L = 95\%$ genome-wide LOD threshold
(compare single-QTL models to the null model)

Choose $\delta = 2 L / \log_{10} n$

With this choice of δ , in the absence of QTLs, we'll include at least one **extraneous** locus, 5% of the time.

Note that now we have

$$\text{BIC}_{\delta} = \log_{10} \text{RSS} + \text{no. markers} \times \left(\frac{2L}{n} \right)$$

Search model space

- Consider the case of additive QTL models, with 100 putative QTLs.
- There are $2^{100} \approx 10^{30}$ possible models, far more than can be inspected individually.
- Need a way to search through this space, to find the good ones.
- This is really a matter of “grunt work.” (More is better; the tradeoff is with computational time.)

Methods of model search

- Forward selection
 - Find the best single-QTL model: q_1^* .
 - Find the best two-QTL model that includes q_1^* : (q_1^*, q_2^*) .
 - Find the best three-QTL model that includes q_1^*, q_2^* : (q_1^*, q_2^*, q_3^*) .
 - Etc.
- Backward elimination
- Forward selection followed by backward elimination
- Stepwise selection
- Randomized algorithms (e.g., MCMC, genetic algorithms, etc.)

Assess performance

- Selection of a model includes two types of errors:
 - Miss important terms (QTLs or interactions)
 - Include extraneous terms
- Unlike in hypothesis testing, we can make **both errors** at the same time!
- **Identify as many correct terms as possible, while controlling the rate of inclusion of extraneous terms.**
- You **can't know** the performance of your procedure with your data—you need to know the truth.
- You **can know**:
 - How a particular procedure performs in simulated cases
 - How a procedure performs in simulated data close to what you've inferred

Science isn't really model selection

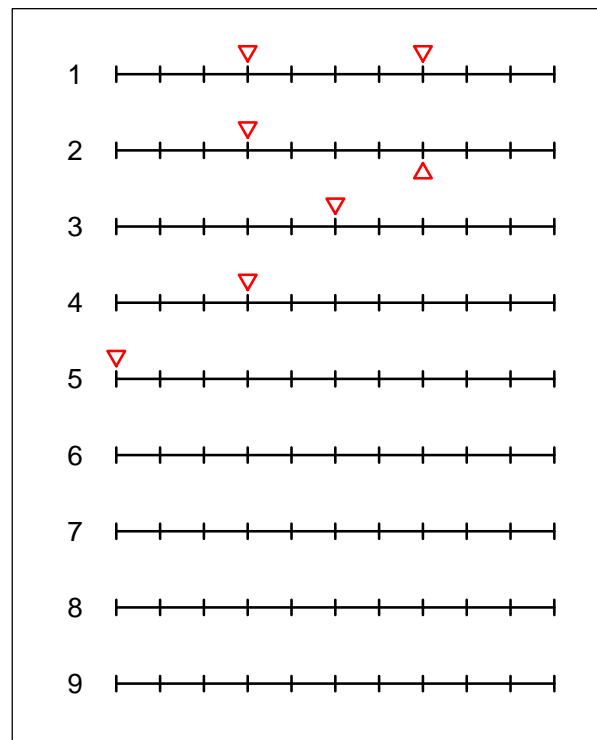
I've said here: pick a good model

Really:

- Want to guide future experiments
- Want some understanding of the **uncertainty** in different aspects of the chosen model

A simulation study

- Backcross with $n=250$
- No crossover interference
- 9 chr, each 100 cM
- Markers at 10 cM spacing; complete genotype data
- 7 QTLs
 - One pair in **coupling**
 - One pair in **repulsion**
 - Three unlinked QTLs
- **Heritability** = 50%
- 2000 simulation replicates



Methods

- ANOVA at marker loci
- Composite interval mapping (CIM)
- Forward selection with permutation tests
- Forward selection with BIC_{δ}
- Backward elimination with BIC_{δ}
- FS followed by BE with BIC_{δ}
- MCMC with BIC_{δ}

→ A **selected marker** is deemed **correct** if it is within 10 cM of a QTL (i.e., correct or adjacent)

A simplified version of CIM

Select a set of markers, S

(e.g., by FS to a fixed number)

For each marker, x , in the genome:

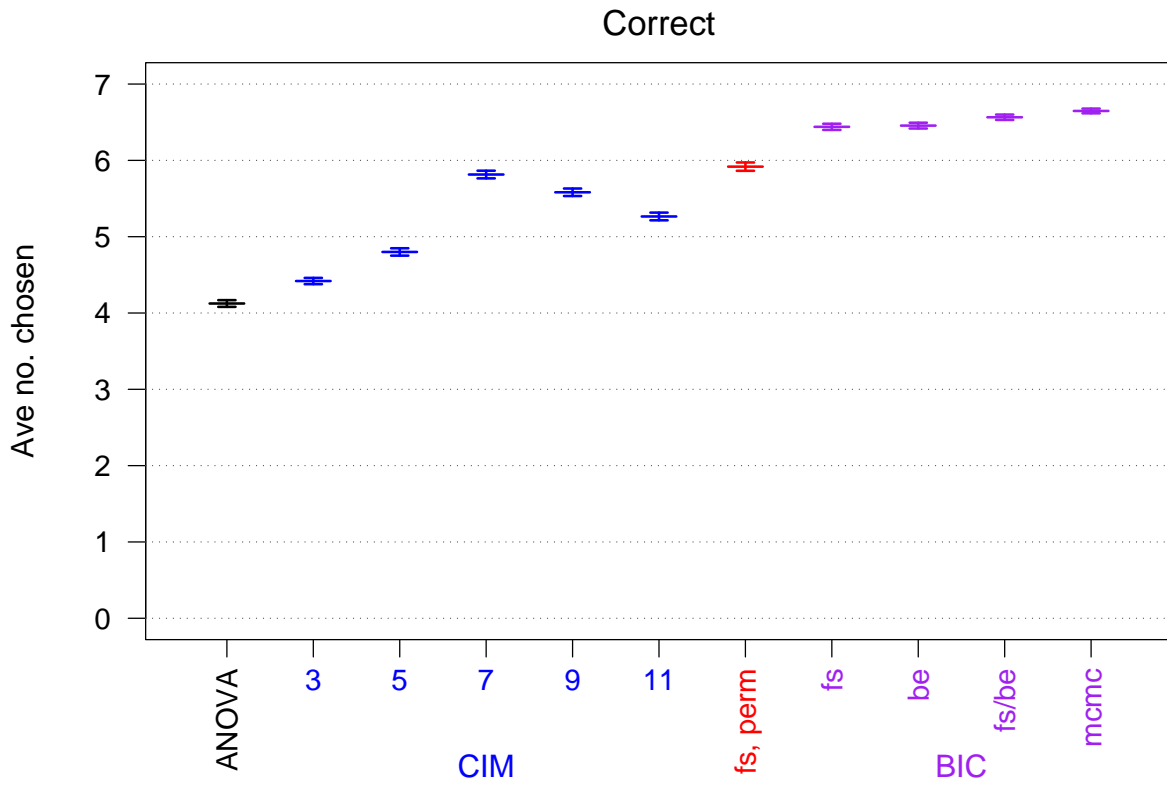
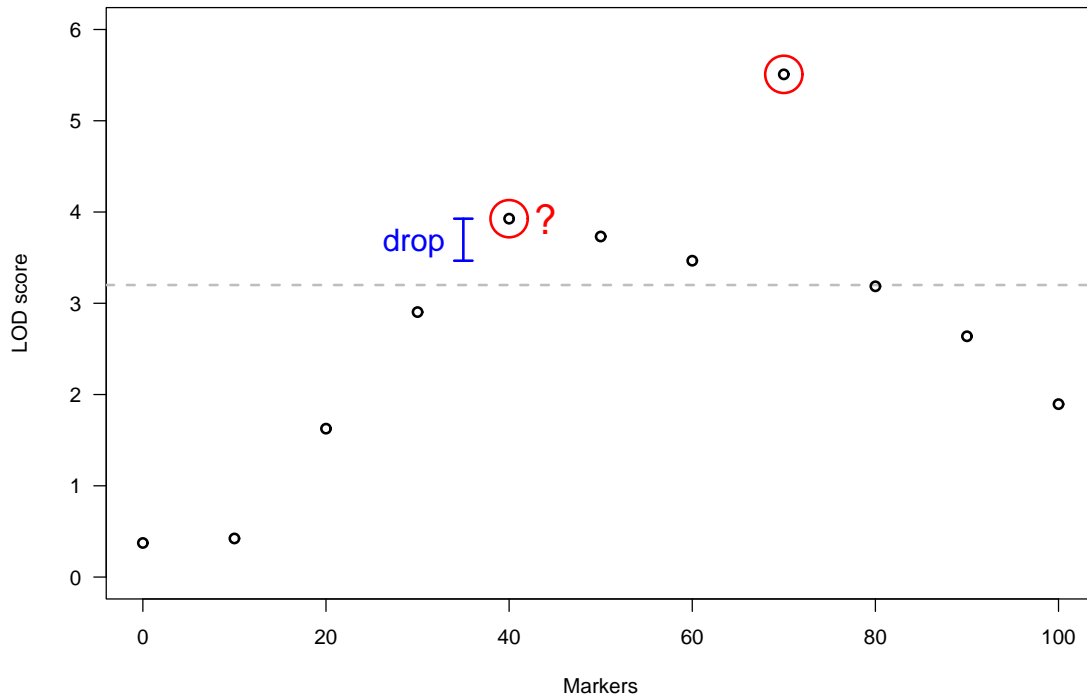
(a) If $x \notin S$, calculate $LOD(x | S)$

(b) If $x \in S$, calculate $LOD(x | S \setminus \{x\})$

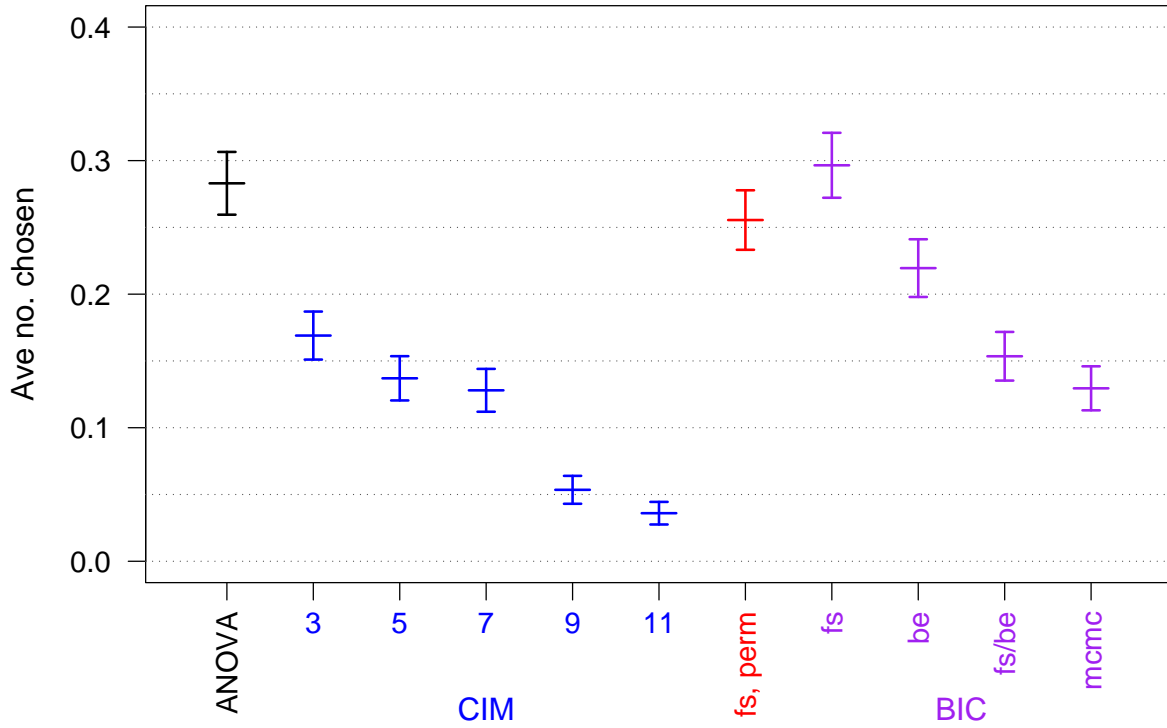
Compare to a genome-wide threshold.

(Take into account the choice of S .)

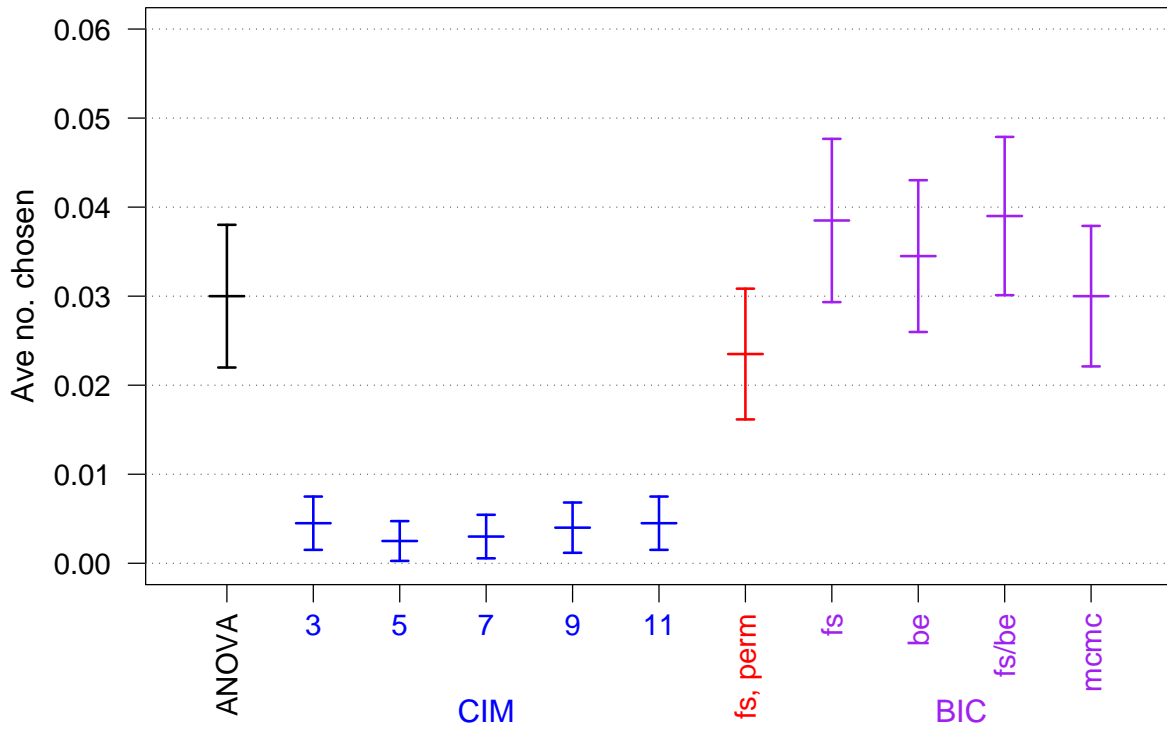
IM / CIM \rightarrow model



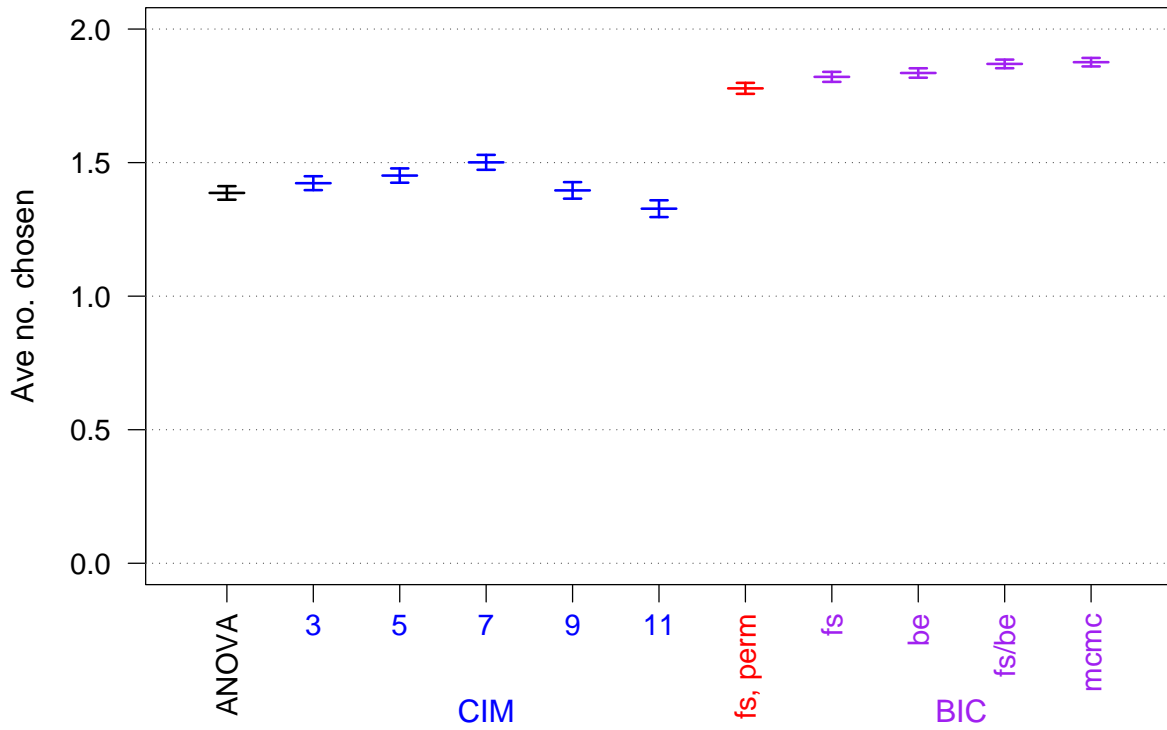
Extraneous linked



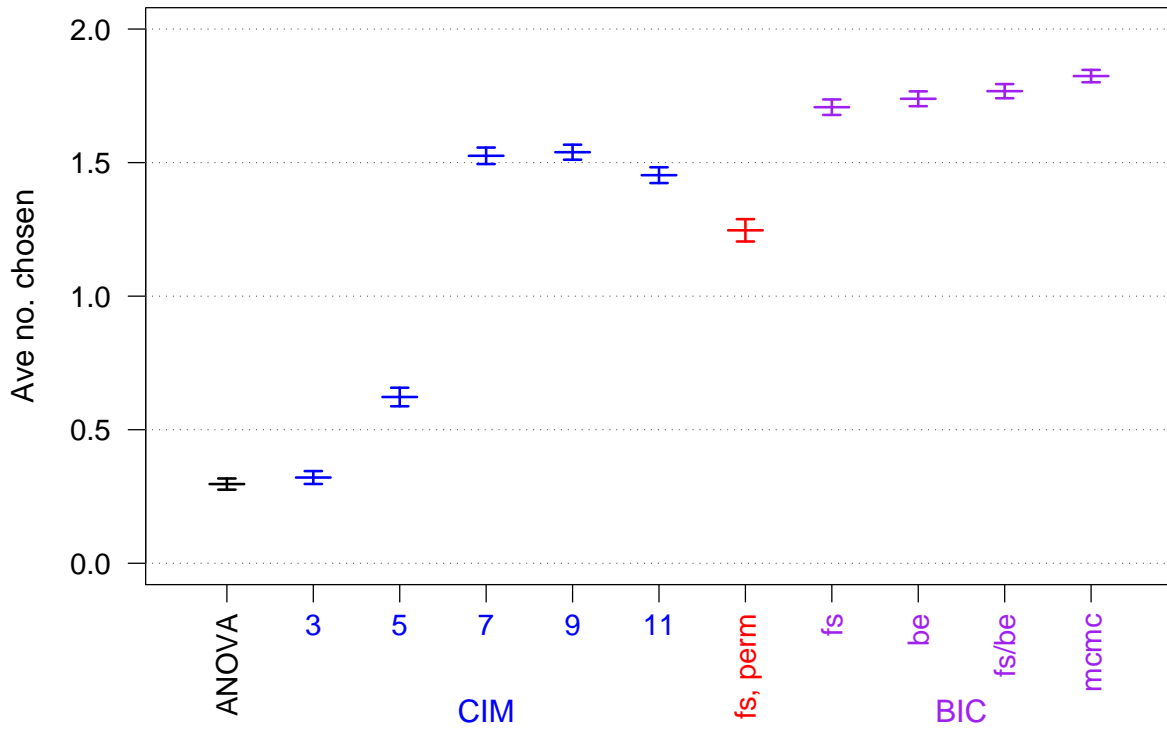
Extraneous unlinked

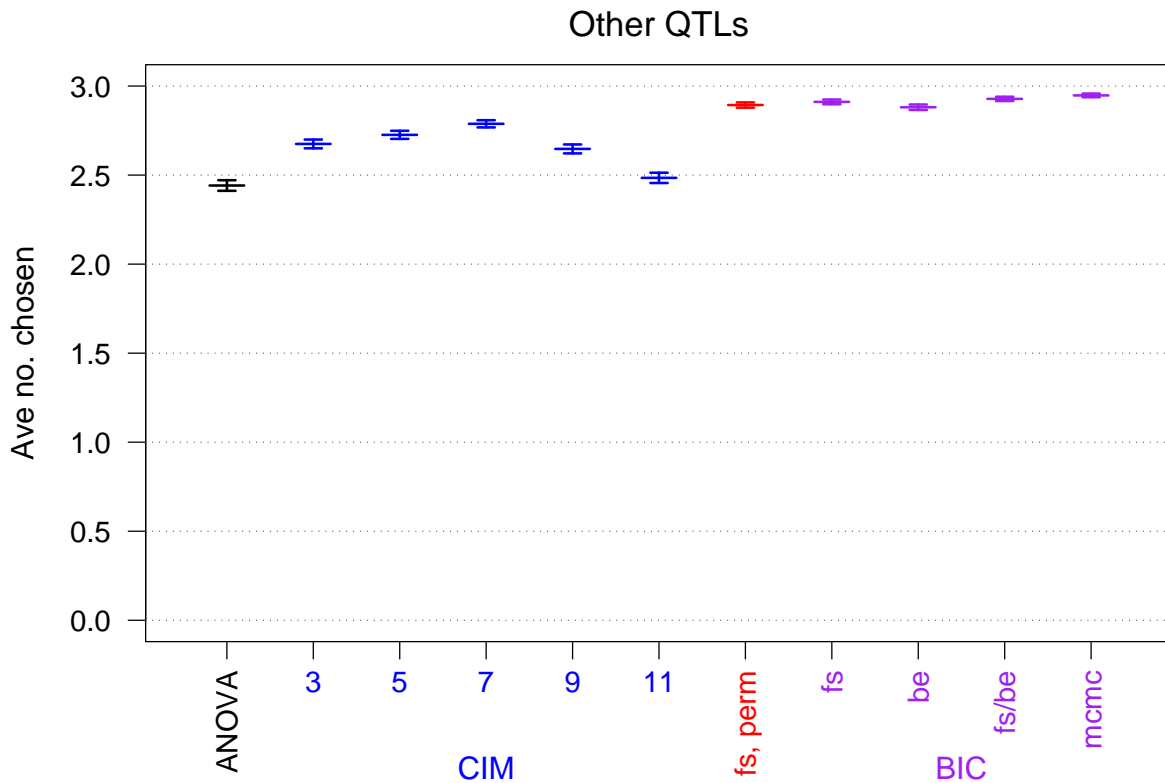


QTLs linked in coupling



QTLs linked in repulsion





Summary

- QTL mapping is a **model selection problem** (rather than **hypothesis testing**).
- Model selection =
 - Select a class of models
 - Select a criterion for comparing models
 - Select a method of searching model space
 - Figure out how your procedure performs
- Key issue: **the comparison of models**.
- Large-scale computer simulations are necessary for assessing the performance of procedures

References

- Broman KW, Speed TP (2002) A model selection approach for the identification of quantitative trait loci in experimental crosses (with discussion). *J Roy Stat Soc B* 64:641–656, 731–775

[Contains the simulation study described above.](#)

- Zeng ZB, Kao CH, Basten CJ (1999) Estimating the genetic architecture of quantitative traits. *Genetical Research* 74: 279–289

[Another paper on the model selection aspects of QTL mapping.](#)

- Miller AJ (2002) *Subset selection in regression*, 2nd edition. Chapman & Hall, New York

[A good book on model selection in regression.](#)